(Sodium Bisulfite)



# **BIONETICS**

SUMMARY OF MUTAGENICITY SCREENING STUDIES CONTRACT FDA 71-268 COMPOUND FDA 71-20 71-268 SODIUM BISULFITE HOST-MEDIATED ASSAY CYTOGENETICS DOMINANT LETHAL ASSAY

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November 24, 1972

Mr. Leonard Appleby, Contracting Officer Department of Health, Education and Welfare Public Health Service Food and Drug Administration, CA-212 5600 Fishers Lane, Room 5C-13 Rockville, Maryland 20852

Reference: Contract FDA 71-268; LBI Project #2311

Dear Mr. Appleby:

Litton Biometics, Inc. is pleased to submit a report for the referenced contract entitled "Mutagenicity Screening Studies" for compound FDA 71-20, Sodium Bisulfite.

Included in this report are the results and raw data of the three tests conducted: Host-Mediated Assay, Cytogenetic Studies; and Dominant Lethal Assay. Eight (8) copies are being submitted for your review.

If there are any questions concerning this report, or, if additional information is required, please do not hesitate to contact us.

Sincerely yours,

LITTON BIONETICS, INC.

OPAF:11s Enclosures (8)

Principal Investigator

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#### I. REPORT

### A. Introduction

Litton Bionetics, Inc. (LBI) has investigated the possible mutagenicity of compounds selected and provided by the Food and Drug Administration under Contract 71-268. LBI's investigation utilized the three mammalian test systems herein described -- Host-Mediated Assay, Cytogenetic Studies and Dominant Lethal Assay. These tests provide information as to the types of genetic damage caused by environmental compounds -- pesticides, chemicals, food additives, drugs and cosmetics.

The Host-Mediated Assay is based upon the assumption that the action of a mutagen on the genetics of bacteria is similar to that in man.

This is further strengthened by the use of an eukaryotic organism (Saccharomyces cerevisiae). Since the mutation frequencies are well established for the indicator organism, any deviation due to the action of the test compound is readily detectable. As some compounds are mutagenic in bacteria and not in the host animal, and vice versa, this test is able to differentiate an action which may have been due to hosts' ability to detoxify or potentiate a suspected mutagen. This action is dependent upon the ability of the compound to gain access to the peritoneal cavity. Coupled with the direct action of the compound on the indicator organism in vitro, the assay provides a clear insight into host-mediation of mutagenicity.

Cytogenetics provides a valuable tool for the direct observation of chromosomal damage in somatic cells. Alteration of the chromosome number and/or form in somatic cells may be an index of mutation. These studies utilized examination of bone marrow cells arrested in C-metaphase from rats exposed to the test compound as compared to positive and negative control animals. If mutational



changes occur, the types of damage expected due to the action of chemicals are structural rearrangements, breaks and other forms of damage to the chromosomal complement of the cells exposed.

For the in vitro cytogenetic studies, we have a more rapid and inexpensive means of determining chronosoma? damage. This is accomplished by observing cells in anaphase. As the chromatids separate and move along the spindle, aberrations may occur. Chromatids which do not migrate to the daughter cells may lead to uneven distribution of parts or of entire chromatids (mitotic nondysjunction). These give rise to "side arm" bridges which have been interpreted as point stickiness or localized failures of chromosome dup?ication point errors. These aberrations (bridges, pseudochiasmata, multipolar cells, acentric fragments, etc.) are extremely sensitive indicators of genetic damage.

The Dominant Lethal Test is an accurate and sensitive measure of the amount and type of fetal wastage which may occur following administration of a potential mutagen. Dominant lethal mutations are indicators of lethal genetic lesions. The effects of mutagens on the chromosomal complement of the spermatozoa of treated males results in alterations of form and number of chromosomes. Structural rearrangements and aneuploidy may lead to the production of non-viable zygotes, early and late fetal deaths, abortions and congenital malformations. In addition, aberrations could lead to sterility or reduced reproductive capacity of the  ${\sf F}_{\vec{1}}$  generation. The action of a mutagen on specific portions of spermatogenesis is also apparent in this test.

### B. <u>Objective</u>

The purpose of these studies is to determine any mutagenic effect of the test compound by employing the Host-Mediated Assay, Cytogenetic Studies



and the Dominant Lethal Assay, both  $\underline{in\ vivo}$  and  $\underline{in\ vitro}$  tests are employed with the cytogenetic and microbial test systems. These tests and their descriptions are referenced in the Appendices A through F.

#### C. Compound

#### Test Material

Compound FDA 71-20, Sodium Bisulfite, Lot #439 1338, as supplied by the Food and Drug Administration.

#### Dosages

The animals employed, the determination of the dosage levels and the route of administration are contained in the technical discussion.

The dosage levels employed for compound FDA 71-20 are as follows for the Cytogenetic Studies <u>in vivo</u> in rats.

Low Level	1.5 mg/kg
Intermediate Level	15.0 mg/kg
LD <sub>5</sub>	150.0 mg/kg
Megative Control	Saline
Positive Control (TEM*)	0.3 mg/kg

The dosage levels employed for compound FDA 71-20 are as follows for the Host-Mediated Assay  $\underline{in}$   $\underline{vivo}$  in mice.

Low. Level	1.5 mg/kg
Intermediate Level	15.0 mg/kg
LD <sub>5</sub>	150.0 mg/kg
Negative Control	Saline
Positive Control (EMS**)	350 mg/kg
(Di@(***)	1 <b>0</b> 0 mg/kg

\* Triethylene Melamine

\*\* Ethyl Methane Sulfonate

\*\*\* Dimethyl Nitrosamine



The dosage levels employed for compound FDA 71-20 are as follows for the Dominant Lethal Assay  $\underline{in}$   $\underline{vivo}$  in rats.

Low Level	1.5 mg/kg
Intermediate Level	15.0 mg/kg
LO <sub>5</sub>	150.0 mg/kg
Negative Control	Saline
Positive Control (TEM*)	0.5 mg/kg

The in vitro Cytogenetic Studies were performed employ-

ing three logarithmic dose levels.

Low Level	2 mcg/m1
Medium Level	20 mcg/m1
High Level	200 mcg/m1
Negative Control	Saline
Positive Control (TEM*)	0.1 mcg/ml

### \*Triethylene Melamine

The discussion of this test is contained in the technical discussion.

### D. <u>Methods</u>

The protocols employed are explained in Appendices C and D.

### E. <u>Summary</u>

Host-Mediated Assay

This compound was non-mutagenic at all dose levels studied.

Cytogenetics

### a. <u>In vivo</u>

The compound produced no detectable significant aberration of the bone marrow metaphase chromosomes of rats when administered orally at the dosage levels employed in this study.

### b. <u>In vitro</u>

The compound produced no significant aberration in the anaphase chromosomes of human tissue culture cells when tested at the desage levels employed in this study.



### Dominant Lethal Study

Compound FDA 71-20 is considered to be non-mutagenic in the Dominant Lethal Study in rats employing the dosage levels used in this study.

### F. Results and Discussion

### Toxicity

### a. In vivo

Compound FDA 71-20 was suspended in 0.85% saline and administered to ten male rats by intubation. The average weight of the animals was 340 grams and each received a dose of 5000 mg/kg. Seven animals died on day 1 and three on day 2. Mecropsy findings indicated a very distended stomach with blood and a whitish, foamy-like substance in the intestine.

Dose levels of 10, 100, 200, 500, 1000, and 2000 mg/kg were selected to determine an acute  $\rm LD_{50}$ . The toxicity data is presented on the  $\rm LD_{50}$  reporting form using the Litchfield-Wilcoxson method.

The LD $_{50}$  was determined as 665 mg/kg. The LD $_{5}$  dose level was derived from the raw data LD $_{50}$  probit line (uncorrected). The LD $_{50}$ 's determined from both corrected probit line and uncorrected probit line were within confidence limits of each other. The acute doses used were LD $_{5}$  - 150 mg/kg; intermediate - 15 mg/kg; and low - 1.5 mg/kg. The subacute dose levels used were the same as those for the acute. The data on the dose levels, numbers of animals and necropsy findings are presented in the toxicity data sheets.

### b. In vitro

The compound was suspended in 0.85% saline at the concentrations listed. It was introduced into tubes containing WI-38 cells in



a logarithmic phase of growth. The cells were observed for cytopathic effect (CPE) and the presence of mitoses at 24 and 48 hours.

Tube No.	No. of Cells	Conc. mcg/ml	CPE	<u>Mi toses</u>
1	5 X 10 <sup>5</sup>	1000	+	+
2		1000	+	+
3		500	+	+
4	н	500	+	+
5	II	100	+	+
6	•	100	+	+
7		10	-	+
8	•	10	· -	+
9		1	-	+
10	н	1	-	+

Since an inhibition of mitoses was observed, a closer range of concentrations was employed, as follows.

1	5 X 10 <sup>5</sup>	500	-	-
2	•	500	<u>+</u>	-
3	•	400	-	-
4		400	-	-
5	ú	300	-	<u>+</u>
6	•	300	-	-
7	ů.	200	-	+
8	, u	200	-	+
9	II	100	-	+
10	H	100	_	+

The 200 mcg/ml concentration was used as the high level. The intermediate level was 20 mcg/ml and the low level was 2 mcg/ml.



CONTRACT FDA 71-268
COMPOUND FDA 71-20
SODIUM BISULFITE



### TOXICITY DATA

CONTRACT FD/: 71-268

COMPOUND FD/. 71-20

### SODIUM BISULFITE

Solvent: 0.85% saline suspension

Animals: Male rats with an average body weight of 340 grams. All animals

were observed for 10 days.

### Range Finding:

	Dose mg/kg	# Dead # Animals	Day of Death and Necropsy
	5000	10/10	Day 1 (7), day 2 (3), distended stomach with blood and foamy substance in the intestine.
LD <sub>50</sub> :			
	- 10	0/5	None
	100	0/5	None
	200	1/5	Day 3, distended stomach with blood and foamy substance in the intestine.
	500	1/5	Day 3, distended stomach with blood and foamy substance in the intestine.
	1000	3/5	Day 1, distended stomach with blood and foamy substance in the intestine.
	2000	5/5	Day 1, distended stomach with blood and foamy substance in the intestine.

## LD50 REPORTING FORM USING LITCHFIELD-WILCOXON METHOD:

DOSE EFFECT CURVE FOR FDA 71-20 Sodium Bisulfite

<u></u>	# dead/ # tested	OBSERVED PERCENT	EXPECTED PERCENT	OES-EXPT PERCENT	CONTRIE. TO (chi)2
<u>100</u> 100	PROPORTION .5 / 5	. 100	.058	.042	.161
200	1 / 5	.200	.156	.044	074
500	1 / 5		.394	194	.788
1000	1 3/5	600	614	014	004
2000	4.5 / 5	.900	.802		.302
	i		<u></u>	<u> </u>	<u> </u>

Total animals =  $\frac{25}{100}$ Number Doses,  $K = \frac{5}{100}$ Animals/Dose =  $\frac{5}{100}$ (CHI)  $\frac{2}{100}$  for n of  $k-2=\frac{7.81}{100}$ 

OBS-EXPT 392 392 (CHI) 2 = 1.329

Degrees of Freedom, n=k-2= 3

since 1.329 is less than 7.81, therefore data not significantly heterogeneous

LD<sub>84</sub> = 2200

LDS0 = 665

LD16 = 205

 $fLD_{50} = S = \frac{2.77}{\sqrt{N'}} = \frac{3.2761}{\sqrt{N'}} = \frac{2.77}{\sqrt{N'}} = \frac{2.77}{\sqrt{10}} = \frac{2.828}{\sqrt{10}}$ 

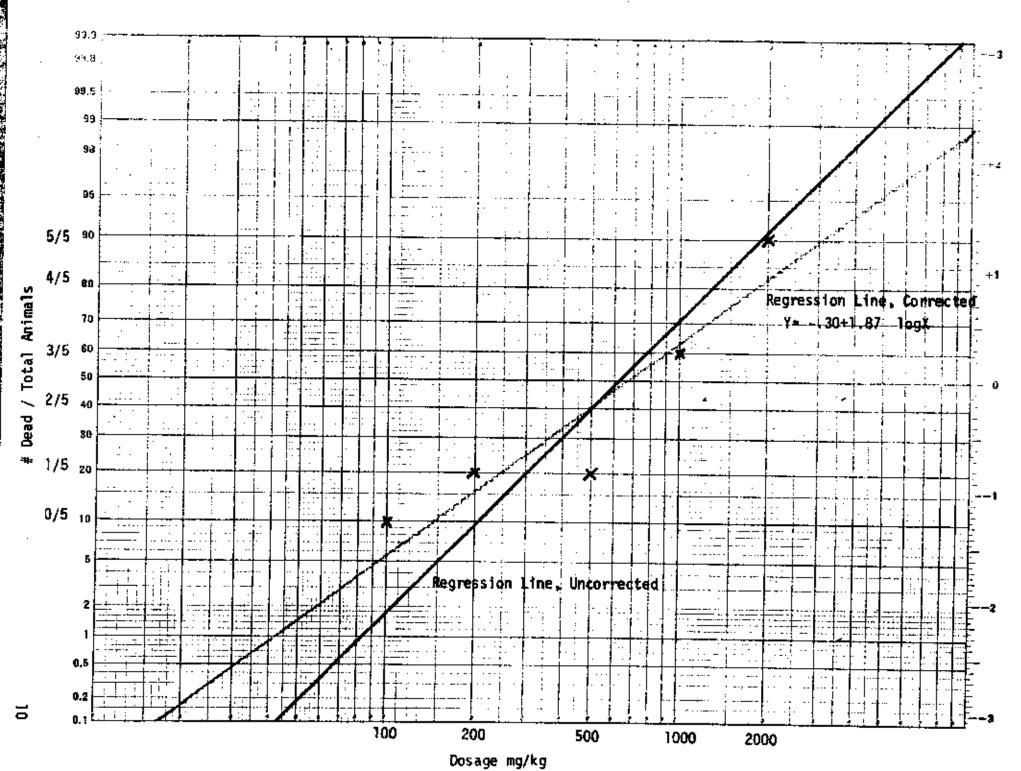
LD<sub>50</sub> x feD<sub>50</sub> = 1880.6

LD50 = 235.1

fLD<sub>50</sub>

LD50 and 19/20 Confidence Limits = P(235 1D 1881) = 95

Attached should be a plot of the dose-effect curve on log-probit paper.



### Host-Mediated Assay - Test I

Compound FDA 71-20 showed no increase in mutation frequencies when tested in vivo against <u>Salmonella</u> TA-1530 and G-46. No increase in recombinant frequencies were seen in tests against <u>Saccharomyces</u> D3.

<u>In vitro</u> studies were negative when either <u>Salmonella</u>

TA-1530 or G-46 were used. Studies with <u>Saccharomyces</u> D3 showed an increased recombinant frequency.



TEST I

CONTRACT FDA 71-268

COMPOUND FDA 71-20

SODIUM BISULFITE



### HOST MEDIATED ASSAY

		· · · · · · · · · · · · · · · · · · ·	SUMMA	RY SHEET			····	
		<del></del>	OUTLIER	S REMOVED				
				EST I			•	
	. COMPOUND: FO	A 71-20	,,,	E31 1				
	** ************************************	/A /15-40	SAL MO	NELLA		Escollaballa		· ·- ·
·	<u>. 14</u>	TA153	10	G-46	•	SACCHAROM	CED 0=3	
		MMF (X 10E-8)	MFT/MFC	MMF (X 10E-8)	MFT/MFC	MRF (X 10E-5)	MRT/MRC	
	ACUTE		- ··· - ·	•				
	NC	3.50		2-14		7.58		
<del></del>	- PC	21.48			40.10	- 36.82	4.86	
	AL Ai .	2.96	-85	2 • 63	1.23	9.72	1.28	
	ALD5	2.62 1.97	.75 	2+21	1.03	2.23	•29	
	<b>-</b> · -	2171		···· 5+49	2•57 <sub></sub>	5.94 ···	.78	
	SUBACUTE							
·				2•14	<u>-</u>	7.58		
	SL SI	2.73	•78	•81	•38	•77	•10	
	- SL05	1.55	,44 •63	2+17	1.01	6.83	.90	
					•38	4.84	64	<u></u>
			· "	•				•
	IN VITRO	TA1530	6-46	<b>-</b> '	 D <b>=3</b>		·· · · · · · · · · · · · · · · · · · ·	
				K CONC	K SURVIVA	L RX 10E	<b>5</b> ;	
; · · · · · · · · · · · · · · · · ·	NC		- <del></del>		<u> </u>			
•	PC [1]							
		·			<u>.</u> .			

### HOST MEDIATED ASSAY

### SUMMARY SHEET

### OUTLIERS INCLUDED

•			-	•
- 1	•	٠.	•	
•	-	•	•	

	. COMPOUND:FDA	71-20		:91 1			
· -	. Gone ografi i rjak	71-20	SALMON	IELLA		SACCHAROMY	CES D-3
		TA153		G-46	<b>5</b>	,	-,
	-	MMF (X 10E-8)	HFT/MFC	MMF (X 10E-8)	MFT/MFC	MRF (X 10E-5)	MRT/MRC
	ACUTE						
 2	NC PC	3.50 21.48	6,14	2•14 85•81	40.10	7.58 36.82	4.86
········	AL	2.68	•77	2.63		9.72	1.28
	AI	2.62	•75	2.21	1.03	2.98	-39
· · · · · · · · · · · · · · · ·	ALD5	1.97	•56	5.49	2.57	- 5 <b>.9</b> 4	.78
·	SUBACUTE						
	NC	3.50	·	2.14		7.58	
	SL	2.73		•95		1.42	•19
	12	2.20				6.83	•90
	SLD5	2•22	£63		-, •3∌ · · · ·	0.19	∵ • <b>ĕ</b> ₽
	IN VITRO	TA1530	G-46		0-3		
			0	# CONC	# SURVIVAL	R X 108	.5
	NC	·	•			•	
٠	PÇ				•		
,	<u></u>						

			HOST MEDI	ATED ASSAY	•		·	<del> </del>
			SUMMAR	Y SHEET		· 		
	COMPOUND: FOA	71-20	TE	ST I				
·		-	SALMON	ELLA		SACCHAROMYC	re n_1	
		TA1536	0	8-46		an animatorities	54 D-3	
<u> </u>	·	MMF	HFT/MFC	MMF	UEB.AUGO			
		(X 10E-8)		(x 10E-8)	well vubiciam, w	(x 10E-5)	MH LAMKE	·•
<del>(4.534-</del> )	- ACUTE					*F #0E-3*		
(10년 2년 1년 - 1년 1년 - 1년	NC	3.50	:	2-14			<del></del>	<del></del>
	P¢	21.48	5.14	85.81	#0.10	8.11 57.0a	, di Mariah	
	AI	2.68		2-65	1.25		7.04 - 1.17	
	AHTD	2.62 1.97	•75	2.21	1.03	5.92	.48	
· · · · · · · · · · · · · · · · · · ·	· · · · · · · · · · · · · · · · · · ·			5.49	2.57	6.55	.81	
·	SUBACUTE		•					··
······································	NC -SU	3.50		2-14	•.	8.11		
	\$1	2.73 2.20	<del></del>	···· <u>95</u>		····· 1.54 ·- ···		·
	SLD5	2.22	•63 •63	2-17	1.01	6.11	.75	
· · · · · · · · · · · · · · · · · · ·	· — · · · · · · · · · · · · · · · · ·		· · · ·		.38	7.74	-95	
	•							·
<del>'</del>	IN VITRO	TA1530	646					
	<b>T</b>	, <b></b>	V40	N CONC			·	
				10	75 75			
· · · · · · · · · · · · · · · · · · ·	TCPD	-	-	10	/3	28		
	HC		··	10	100 68	28 6		

## HOST MEDIATED ASSAY (OUTLIERS REMOVED)

### SUMMARY SHEET

COMPOUND: FDA	71-20	TE	ST I			
	TA152	SALMON			SACCHAROMYO	ES D+3
	TA153	241	G-46	)		
	NMF (X 10E-8)	MFT/MFC	MMF (X loe-8)	MFT/MFC	MRF (X 10E-5)	MRT/MRC
ACUTE NC PC - AU A1 ALD5	3.50 21.48 2.96 2.62 1.97	6.14 .85 .75	2.14 85.81 2.63 2.21 5.49	40.10 1.23 1.03 2.57	8.11 57.08 9.46 1.57 6.55	7.04 1.17 .19
SUBACUTE NC Su SI SLD5	3.50 2.73 1.55 2.22	.78 .44 .63	2.14 .81 2.17 .81	.38 1.01 .38	8.11 .32 6.11 4.96	.04 .75 .61
IN VITRO	TA1530	G-46	% CONC	D-3 % SURVIVAL	R X 10E5	
NC SAM PC	E AS ON SUMMARY	SHEET PRECEDI	NG			
CSCX CSC85F 24 NOV 72 CARDS IN 73 OUT 0	15:20:19 USE LINES 47	ER CFU007 7 PROCESSI	190 IG TIMT	2.85 SECONDS		

b. HOST-MEDIATED ASSAY DATA SHEETS

TEST I

CONTRACT FDA 71-268

COMPOUND FDA 71-20

SODIUM BISULFITE



### TEST I

		1 <b>£</b> 21 1		_
COMPOUND:	FDA 71-20		ORGANISM: SAL	MONELLA TA1530
DOSE, LEVE	LE NEGATIVE COM	TROL - WATER		
TREATMENT	: IN VIVO, ORAL	., ACUTE	DATE STARTED:	MARCH 10, 197
	A	B	C TOTAL NO	D
ANIMAL NUMBER	RAW CFU X 10E7/0.6ML	TOTAL CFU X 10E8/1.0ML	TOTAL NO. MUTANTS X 10E0/1.0ML	MUTATION FRE (C/B) X 10E-8
1	36.10	6.02	14.00	2.33
2	24+20	4.03	13.00	3.22
3	33.10	5.52	5.00	•91
4.	37.20	6.20	6.00	•97
5	12.00	2.00	15.00	7.50
2 3 4 5 6 7	<b>3</b> 0•10 9•00	5.02 1.50	8,00 12,00	1.59 8.00
NO. OF AN	NIMALS EQUALS	7 EQUALS 3		
		COL. B	COL. C	COL. D
		(X 10E8)	(X 10E0)	(X 10E-8)
	MEAN	4,33	10-43	3.50
• .	RANGE	4.70	10.00	7.09 8.00
	MAX	6.20	15.00 5.00	•91
NO OUTLIE	MIN ERS	1.50	5400	4 7 <u>†</u>

SCX CSC85F 21 NOV 72 17:20:13 USER CFU007 200 ARDS IN 236 OUT 0 LINES 63 PROCESSING TIME 5.97 SECONDS

#### TEST I

COMPOUND: FDA 71-20

ORGANISM: SALMONELLA TA1530

DOSE LEVEL: POSITIVE CONTROL - DMH - 100 MG/KG

TREATMENT: IN VIVO, ORAL, ACUTE DATE STARTED: MARCH 10, 1972

	A	B	C TOTAL NO.	D MUTATION
ANIMAL	RAW CFU X	TOTAL CFU X	MUTANTS X	FRE (C/B)
NUMBER	10E7/0.6ML	10E8/1.0ML	10E0/1.0ML	X 10E-8
1	30.00	5.00	72.00	14.40
	12.90	2.15	24.00	11.16
2 3	15.20	2.53	109.00	43.03
	33.10	5.52	112.00	20.30
ξ.	57.60	9.60	22.00	2.29
2	12.00	2.00	72.00	36+00
4 5 6 7	15.90	2.65	32.00	12+08
8	19.50	3.25	106.00	32.61
	IIMALS EQUALS	8 EQUALS 2		•
TOTAL CEC	OUT OF HANDE	EACHED =		
		COL - B	CDI - C	COL. D

		COL. B	CDL. C	COL. D
		(X 10EB)	(X 10E0)	(X 10E-8)
	MEAN	4.09	68.62	21.48
	RANGE	7.60	90.00	40.73
	MAX	9.60	112.00	43.03
	MIN	2.00	22.00	2+29
NO OUTLIERS			1 16 3	grand Article (1986)

CSCX CSC85F 21 NOV 72 17:20:34 USER CFU007 200

ARDS IN 236 OUT 0 LINES 64 PROCESSING TIME 5.87 SECONDS

;			TEST I		
,	COMPOUND:	FDA 71-20		ORGANISM: SAL	ONELLA TA1530
i	DOSE LEVEL	: LOW - 1.5 MG/	<b>/</b> KG		
		IN VIVO, ORAL		DATE STARTED:	MARCH 10: 1972
		A	₿	C, NO.	D MUTATION
		BAN CELL V	TOTAL CFU X	TOTAL NO. MUTANTS X	FRE (C/B)
	ANIMAL	RAW CFU X 10E7/0.6ML	10E8/1.0ML	10E0/1.0ML	X 10E-8
•	NUMBER	TOCAS DECME			3 70
}	1	16.20	2.70	10.00	3.70 3.31
	2	14.50	2.42	8.00	2.55
7	3	16.50	2.75	7.00	1.63
J.	4	36.80	6.13	10.00	4.62
	5	13.00	2.17	10.00	.20
•	2 3 4 5 6 7	29.50	4.92 6.63	22.00	3.32
_	7	39.80	2.80	6.00	2.14
•	<b>B</b> .	16.B0	2.40	4.00	1.67
•	9	14.40 16.30	2.72	10.00	3.68
	10	10+30	_ <del> , _</del>		
	NO. OF AN	IMALS EQUALS	10		
7			COL. B	COL. C	coL, D
			(X 10E8)	(X 10E0)	(X 10E-8)
		MEAN	3.56	8.80	2.68
7		RANGE	4.47	21.00	4.41
1		MAX	6,63	22.00	4.62
•		MIN	2,17	1.00	+20 <sub>.</sub>
]					
j		*	SUMMARY WITH	OUTLIERS REMOV	Ep`
ሳ			•		
1			COL. B	COL. C	ÇOL, D
•			(X 10E8)	(X 10E0)	(X 10E-8)
13		MEAN	3,41	9.67	2.96
ŀ		RANGE	4.47	18.00	2,98
•		MAX	6.63	22.00	4,62
		MIN	2.17	4.00	1.63
}		F1 <b>p</b> 17	-,-,-		
	SC85F 21 NO	v 72 17:20:53	USER CFU007	200	
CSCX C	arebi ST Mo	A 15 Triboton			

CARDS IN 236 OUT 0 LINES 76 PROCESSING TIME

7.15 SECONDS

ř			TEST I		
רן	COMPOUND: F	DA 71-20	1231 1	ORGANISM: SAL	MONELLA TA1530
, j	DOSE LEVEL:	INTERMEDIAT	E - 15 MG/KG	,	
7	TREATMENT:	ÍN VIVO, ORA	L. ACUTE	DATE STARTED:	MARCH 10, 1972
7		A	. 8	с	D
	ANIMAL NUMBER	RAW CFU X 10E7/0.6ML	TOTAL CFU X 10E8/1.0ML	TOTAL NO. MUTANTS X 10E0/1.0ML	MUTATION FRE (C/B) X 10E-8
با ب	1 2 3	13.80 7.00 18.90	2.30 1.17 3.15	6.00 4,00 2.00	2.61 3.43
** <b>!</b>	4 5 6 7	24.00 22.70 23.90	4.00 3.78 3.98	7.00 9.00 6.00	.63 1.75 2.38 1.51
*	, 8 9 10	12.10 30.00 12.50 22.80	2.02 5.00 2.08 3.80	9,00 11.00 8.00	4.46 2.20 3.84
' . ፲	NO. OF ANIMA		10	13.01	3.42
]		MEAN RANGE	COL. B (X 1058) 3.13 3.83	COL. C (X 10E0) 7.50 11.01	COL. D (X 10E-8) 2.62 3.83
]	NO OUTLIERS	MAX Min	5,00 1,17	13.01 2.00	4.46 .63
CSCX CSC8	5F 21 NOV 72	17:21:21	USER CFU007	200	
CARDS IN	236 OUT.	0 LINES	65 PROCESSING	TIME 6.87	SECONDS

: GNUO PMO	FDA 71-20	TEST I	ORGANISM: SALM	IONELLA TA1530
DOSE LEVEL	: LD5 - 150 MG	e/KG		
TREATMENT	IN VIVO+ ORAL	. ACUTE	DATE STARTED:	MARCH 10. 197
	A	. <b>B</b>	C.	ð
			TOTAL NO.	MUTATION FRE (C/B)
ANIMAL	RAW CFU X	TOTAL CFU X	MUTANTS X 10E0/1.0ML	X 10E-8
NUMBER	10E7/0.6ML	10E8/1.0ML	TOPOSTAGAL	Y, 105,-0
1	39.00	6.50	9.00	1.38
2	12.20	2.03	6.00	2.95
3	12.60	2.10	4.00	1.90
4	18.10	3.02	2.00	•66
5	15.60	2.60	7.00	2.69
6	21.60	3.60	5.00	1.39 3.63
2 3 4 5 6 7 8	18.20	3.03	11.00 3.00	2.90
8,	6+20	1.03 1.90	2.00	1.05
9 10	11.40 21.80	3.63	4.00	1.10
NO. OF AN	IMALS EQUALS	10		
		COL, B	COL. C	COL. D (X 10E-8)
	****	(X 10E8) 2.95	(X 10E0) 5.30	1.97
	MEAN	2.95 5.47	9.00	2.96
	RANGE. MAX	6.50	11.00	3,63
	MIN .	1.03	2.00	. 66
NO SUTLIE	ŘŠ*********		1 1	

CSCX CSC85F 21 NOV 72 17:21:42 USER CFU007 200

CARDS IN 236 OUT D LINES 65 PROCESSING TIME 6. 3 SECONDS

ព្	СОМРОШНО:	FDA 71-20	TEST I	ORGANICM: CAL	MONELLA TA1530
_	DOSE, LEVEL	.: LOW - 1.5 M	G/KG	Stranger SAL	MUNICIPE TAIS30
	TREATMENT:	IN VIVO, ORA	L. SUBACUTE	DATE STARTED:	MARCH 10, 1972
<b>m</b> :		A	В	C <sub>.</sub>	מ
roj j	ANIMAL NUMBER	RAW CFU X 10E7/0.6ML	TOTAL CFU X 10E8/1.0ML	TOTAL NO. MUTANTS X 10EO/1.0ML	MUTATION FRE (C/B) X 10E-8
	1 2 3	11:20 51:00 7:00	1.87 8.50 1.17	9.00 8.00 2.00	4.82 .94
r <del>s</del>	4 5 6 7	14.70 22.00 17.20	2.45 3.67 2.87	11.00 8.00 7.00	1.71 4.49 2.18
n	9	<b>30.</b> 00 13.20 12.40	5+00 2•20 2+07	9,00 3,00 10,00	2.44 1.80 1.36 4.84
7	NO. OF ANIM Total Cfu (	MALS EQUALS OUT OF RANGE E	9 QUALS 1	,	4.0,4
•		MEAN RANGE	COL. 8 (X 10E8) 3,31	COL. C (X 10E0) 7.44	COL. D (X 10E-8) 2.73
•	NO OUTLIERS	MAX MIN	7.33 8.50 1.17	9.00 11.00 2.00	3.90 4.84 .94
' CSCX	CSC85F 21 NOV 7	2 17;22; 3 <sub>U</sub>	SER CFU007	200	•

CARDS IN 236 OUT 0 LINES 65 PROCESSING TIME 6. 1 SECONDS

T	c	c	Ŧ		T
	C	э	1	-	1

		·	TEST .I		
	COMPOUND	: FDA 71-20		ORGANISM: SALM	ONELLA TAISSU
	DOSE LEV	EL: INTERMEDIATE	- 15 MG/KG		
		T: IN VIVO, ORAL		DATE STARTED:	MARCH 10: 1972
			В	С	D
		A	U	TOTAL NO.	MUTATION
	ANITMAL	RAW CFU X	TOTAL CFU X		FRE (C/B)
	ANIMAL NUMBER	10E7/0.6ML	10E8/1.0ML	10E0/1.0ML	X 10E-8
	•	56+10	9.35	3.00	•32
	1 2	12.60	2.10	8.00	3.81
	2 3 4	7.30	1.22	9.00	7.40
	7	19.20	3.20	7,00	2.19
	5	52.20	8+70	10.00	1.15
	5 6 7	12.20	2.03	2.00	•98 • 71
	7	14.00	2.33	4.00	1.71 1.42
	В	37.90	6.32	9.00	.84
	9	14.30	2.38	2.00	••,•,
	NO. OF	ANIMALS EQUALS DEAD ANIMALS EQU	9 ALS 1		
			COL. B	COL. C	COL. D
			(X 10EB)	(X 10E0)	(X 10E-8)
		MEAN	4.18	6.00	2.20 7.08
		RANGE	8.13	8.00 10.00	7.40
		MAX	9,35	2.00	.32
		MIN	1.22	2000	•,
		•	SUMMARY WITH	OUTLIERS REMOVE	p
			· <b>-</b>	na. · · ·	COL. D
•			COL. B	COL. C	(X 10E-8)
			(X 10E8)	(X 10E0)	1.55
		MEAN	4.55 7.33	5 <b>.63</b> 8 <b>.0</b> 0	3.49
,		RANGE	7.32 9.35	10.00	3,81
1		MAX		2.00	, .32
, i		in saas MAN aara	ALE SEE MARKET	, <u> </u>	•
רכרץ רב	CASE 21 A	10V 72 17;22:25	USER CFUDO7	200	
TOUR OU		reconstruction of the second	· 	==================================	THE COMMUNICATION

76 PROCESSING TIME

0 LINES

CARDS IN 234 OUT

6.31 SECONDS

COMPOUND: FDA 71-20	ORGANISM: SALMONELLA TA1530	
DOSE LEVEL: LD5 - 150 MG/KG		
TREATMENT: IN VIVO, ORAL, SUBACUTE	DATE STARTED: MARCH 10, 1972	
_ ·		

	A	В	c	Đ
ANIMAL NUMBER	RAW CFU X 10E7/0.6ML	TOTAL CFU X 10E8/1.0ML	TOTAL NO. MUTANTS X 10E0/1.0ML	MUTATION FRE (C/B) X 10E-8
1 2 3 4 5 6 7 8	29.00 16.10 20.90 38.80 25.80 17.70 23.90 13.80	4.83 2.68 3.48 6.47 4.30 2.95 3.98 2.30	10.00 9.00 6.00 4.00 8.00 8.00 9.00	2.07 3.35 1.72 .62 1.86 2.71 1.51 3.91
NO. OF	ANIMALS EQUALS DEAD ANIMALS EQUAL CONTAMINATED EQUAL		•	
NO OUTL	MEAN RANGE MAX MIN IERS	COL. B (X 10E8) 3.88 4.17 6.47 2.30	COL. C (X 10E0) 7.50 6.00 10.00 4.00	COL. D (X 10E-8) 2.22 3.29 3.91 .62

CSCX CSC85F 21 NOV 72 17:22:51 USER CFU007 200

CARDS IN 232 OUT 0 LINES 65 PROCESSING TIME 6.33 SECONDS

### TEST I

COMPOUND: FOA 71-28

ORGANISM: SALMONELLA 6-46

DOSE LEVEL! NEGATIVE CONTROL - WATER

LINES

234 OUT

CARDS IN

TREATMENT: IN VIVO. ORAL, ACUTE

DATE STARTED: FEBRUARY 11: 1972

ANIMAL NUMBER	A RAW CFU X 10E7/0.6ML	TOTAL CFU X	C TOTAL NO. MUTANTS X 10E0/1.0ML	D MUTATION FRE (C/B) X 10E-8
1 2 3 4 5 6 7 8	20.30 24.20 16.30 19.10 18.20 16.60 28.60 24.80	3.38 4.03 2.72 3.18 3.03 2.77 4.77	5.00 14.00 6.00 5.00 6.00 5.00 8.00	1.48 3.47 2.21 1.57 1.98 1.81 1.68 2.90
እነለ ለ <u>ሮ</u> ርብ	IMALS EQUALS NTAMINATED EQU OUT OF RANGE	8 ALS 1 EQUALS 1	,	
	MEAN RANGE MAX MIN	COL. B (X 10E8) 3.50 2.05 4.77 2.72	COL. C (X 10E0) 7.63 9.00 14.00 5.00	COL. D (X 10E-8) 2.14 1.99 3.47 1.48
NO OUTLIE	RS			•
CSCX CSC85F 21 NOV	72 17:17:46	USER CFU007	200	_
CARRE TN 234 OUT	o LINES	65 PROCESSING	TIME	6. 1 SECONDS

		TEST I		
COMPOU ID:	FDA 71-20		ORGANISM: SAL	MONELLA G≕45.
DOSE LEVEL	. POSITIVE CON	NTROL - DMN -		, , , , , , , , , , , , , , , , , , ,
			DATE STARTED:	FEBRUARY 11
	A	В	C.	Ð
ANIMAL NUMBER	RAW CFU X 10E7/0.6ML	TOTAL CFU X 10E8/1.0ML	MUTANTS X 10ED/1.0ML	HUTATION FRE (C/B) X 10E-8
1 2 3	31.20 27.10	5.20 4.52	100.00 120.00	19.23 26.57
4 5	21.10 27.00	3.52 4.50	340.00	15.29 96.68 46.67
7 8	13.30 22.40 22.90	3.73	470.00 480.00	212.03 128.57
NO. OF DEAD	D ANIMALS EQUA	8 (5 1	440400	141.48
NO. OF CON	TAMINATED EQUA	L5 1		
	MEAN Pange	COL. B (X 10E8) 4.09	COL. C (X 10E0) 292.50	COL. D (X 10E-8) 85.81
NO OUTLIERS	MAX Min	3.02 5.23 2.22	460.00 540.00 80.00	196.74 212.03 15.29
	TREATMENT:  ANIMAL NUMBER  1 2 3 4 5 6 7 8 NO. OF ANI NO. OF CON	TREATMENT: IN VIVO. ORAL  A  ANIMAL RAW CFU X NUMBER 10E7/0.6ML  1 31.20 27.10 3 31.40 4 21.16 5 27.00 6 13.30 7 22.46 8 22.90  NO. OF ANIMALS EQUALS NO. OF DEAD ANIMALS EQUA NO. OF CONTAMINATED EQUA  MEAN RANGE MAX MIN	COMPOUND: FDA 71-20  DOSE LEVEL: POSITIVE CONTROL - DMN -  TREATMENT: IN VIVO, ORAL, ACUTE   A B  ANIMAL RAW CFU X TOTAL CFU X NUMBER 10E7/0.6ML 10E8/1.0ML  1 31.20 5.20 2 27.10 4.52 3 31.40 5.23 31.40 5.23 4 21.10 3.52 5 27.00 4.50 5 13.30 2.22 7 22.40 3.73 8 22.90 3.82  NO. OF ANIMALS EQUALS 8 NO. OF DEAD ANIMALS EQUALS 1 NO. OF CONTAMINATED EQUALS 1	COMPOUND: FDA 71-20  DOSE LEVEL: POSITIVE CONTROL - DMN - 100 MG/KG  TREATMENT: IN VIVO. ORAL. ACUTE  A  B  C  ANIMAL RAW CFU X TOTAL CFU X MUTANTS X NUMBER 10E7/0.6ML 10E8/1.0ML 10E0/1.0ML  1 31.20 5.20 100.00 2 27.10 4.52 120.00 3 31.40 5.23 80.00 4 21.16 3.52 340.00 5 27.00 4.50 210.00 5 27.00 4.50 210.00 6 13.30 2.22 470.00 7 22.40 3.73 480.00 7 22.40 3.73 480.00 8 22.90 3.82 540.00  NO. OF ANIMALS EQUALS 8 NO. OF DEAD ANIMALS EQUALS 1 NO. OF CONTAMINATED EQUALS 1

CARDS IN 232 OUT 0 LINES 65 PROCESSING TIME 5.79 SECONDS

#### TEST I

COMPOUND: FDA 71-20

ORGANISM: SALMONELLA G-46

DOSE LEVEL: LOW - 1.5 MG/KG

TREATMENT: IN VIVO, ORAL, ACUTE DATE STARTED: FEBRUARY 11, 1972

	A	В	C TOTAL NO.	D MUTATION
ANIMAL NUMBER	RAW CFU X 10E7/0.6ML	TOTAL CFU X 10E8/1.0ML	MUTANTS X 10E0/1.0ML	FRE (C/8)
1	30+10	5.02	8.00	1.59
2	15,80	2.63	13.00	4.94
3	16.30	2.72	10.00	3.68
4	20.80	3.47	5,00	1.44
5	14.90	2.48	7.00	2.82
6	16.90	2.82	8.00	2.84
7	7.90	1.32	1.00	•76
8	12.20	2.03	6.00	2.95
NO. OF	ANIMALS EQUALS	8	•	

NO. OF DEAD ANIMALS EQUALS

		COL, B	COL. C	ÇOL. D
		(X 10E8)	(X 10E0)	(X 10E-8)
	MEAN	2.81	7.25	2.63
	RANGE	3.70	12.00	4.18
	MAX	5.02	13.00	4.94
	MIN	1.32	1.00	.76
NO OUTLIERS				

NO OUTLIERS

CSCX CSC85F 21 NOV 72 17:18:28 USER CFU007 200

CARDS IN 232 OUT 0 LINES 64 PROCESSING TIME 5.71 SECONDS

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		TEST I		
COMPOUND:	FDA 71-20		ORGANISM: SAL	MONELLA G-46
DOSE LEVE	L: INTERMEDIATI	E - 15 MG/KG		•
TREATMENT	: IN VIVO, ORA	L, ACUTE	DATE STARTED:	FEBRUARY 11,
	A	· B	_ <b>c</b> ,	D
ANIMAL	RAW CFU X	TOTAL CFU X	TOTAL NO. Mutants X	MUTATION FRE (C/B)
NUMBER	10E7/0.6ML	10E8/1.0ML	10E0/1.0ML	X 10E-8
1 .	24.70	4.12	22.00	5.34
2 3 4 5 6 7 8	19.40	3.23	14.00	4.33
3	11.60	1.93	3.00	1.55
4.	19.70	3.28	3.00	•91
5	30.50	5≠08	5,00	•98
<u>, 6</u>	23.00	3.83	6.00	1.57
7	19.10	3.18	6.00	1.88
<b>5</b> .	10.70	1.78	2.00	1.12
NO. OF CO	IMALS EQUALS NTAMINATED EQUA ITH ZERO MUTAN]	8 ALS 1 IS EQUAL 1	•	
		COL. B	COL. C	COL. D
		(X 10EB)	(X 10E0)	(X 19E-8)
	MEAN	3.31	7.63	2.21
	RANGE	3,30	20.00	4.43
	MAX	5,08	22.00	5.34
	MIN,	1.78	2.00	.91
NO OUTLIE	R5 .			-
85F 21 NOV	72 17:18:49	HSER CEHOAT	200	

CSCX CSC85F 21 NOV 72 17:18:49 USER CFU007 200

CARDS IN 232 OUT 0 LINES 65 PROCESSING TIME 5.77 SECONDS

TEST 1

		1E5  I		
СОМРОИМЫ	FDA 71-20		ORGANISM: SAL	MONELLA 6-46
DOSE LEVE	L: LD5 - 150 M	5/KG		
TREATMENT	: IN VIVO, ORAL	. ACUTE	DATE STARTED:	FEBRUARY 11
		<b>B</b>	C	. D
ANIMAL	RAW CFU X	TOTAL CITU X	TOTAL NO.	MUTATION
NUMBER	18E7/0.6ML	10E8/1.0ML	10E0/1.0ML	FRE (C/B) X 10E-8

1.67

1.90

3.08

1.67

1.37

3.48

3.92

NO. OF DEAD ANIMALS EQUALS 1
TOTAL CFU OUT OF RANGE EQUALS 1

10.00

18.50

10.00

20+90

23.50

8.20

6

NO OUTLIERS

11.40:

COL. B COL. C COL. D (X 10E8) (X 10E0) (X 10E-8) MEAN-2.38 12.25 -5.49 RANGE 2.55 23.00 7.55 MAX 3.92 28.00 9.08 MIN-

200

10,00

5.00

14.00

28,00

12.00

15.00

6.00

8.00

CSCX CSC85F 22 NOV 72 21:18:37 USER CFU007

GARDS IN 234 OUT 0 LINES 65 PROCESSING TIME 5.82 SECONDS

1972

5.08

3.00

7.37

9.08

4.80

8.78

4.31

1.53

#### TEST I

COMPOUND: FDA 71-20 ORGANISM: SALMONELLA G-46

DOSE LEVEL: LOW - 1.5 MG/KG

TREATMENT: IN VIVO, ORAL, SUBACUTE DATE STARTED: FEBRUARY 11, 1972

	A	В	C TOTAL NO.	D Mutation	
ANIMAL	RAW CFU X	TOTAL CFU X	MUTANTS X	FRE (C/B)	
NUMBER	10E7/0.6ML	10E8/1.0ML	10E0/1.0ML	X 10E-8	
1	34.10	5.68	3.00	•53	
1 2 3	38.00	6.33	6.00	•95	
3	32+00	5.33	3,00	+56	
4	19.60	3.27	4.00	1.22	
5	22.00	3.67	7,00	1.91	
4 5 6 7	27.90	4.65	4.00	• 66	
7	. 25.90	4.32	4.00	•93	
8,	29.30	4.88	3.00	.61	
NO. OF AN	IMALS EQUALS	8	1		
	OUT OF RANGE	•			
		COL, B	COL. C	COL. D	
		(X 10E8)	(X 10E0)	(X 10E-8)	
	MEAN.	4.77	4.25	•95	
	RANGE	3.07	4.00	1.38	

4.77	4.25	•95
3.07	4.00	1.38
6.33	7.00	1.91
3.27	3.00	•53
	·	

### \* SUMMARY WITH OUTLIERS REMOVED

	COL, B	COL. C	COL. D
	(X 10E8)	(X 10E0)	{X 10E-8}
MEAN	4.92	3.86	.81
RANGE	3.07	3.00	.70
MAX	6.33	6.00	1.22
MIN	3.27	3.00	•53

CSCX CSC85F, 21 NOV 72 17:19: 7 USER CFU007 200

MIN

CARDS IN 236 OUT 0 LINES 75 PROCESSING TIME 6. 1 SECONDS

#### TEST I

COMPOUND: FDA 71-20 ORGANISM: SALMONELLA G-46

DOSE LEVEL: INTERMEDIATE - 15 MG/KG

TREATMENT: IN VIVO, ORAL, SUBACUTE DATE STARTED: FEBRUARY 11, 1972

	A	B	C	D
	man America		TOTAL NO.	MUTATION
ANIMAL	RAW CFU X	TOTAL CFU X	MUTANTS X	FRE (C/B)
NUMBER	10E7/0.6ML	10E8/1.0ML	10E0/1.0ML	X 10E-8
1	49.30	8.22	25.00	3.04
2	38.50	6.42	18.00	2.81
3	15.50	2.58	6.00	2.32
4	30.60	5.10	10.00	1.96
2 3 4 5 6 7 8	11.10	1.85	2.00	1.08
6	26.90	4.48	5.00	1.12
7	7.70	1.28	2.00	1.56
8	47.00	7.83	27.00	3.45
NO. OF AN	IMALS EQUALS	8	1	
	OUT OF RANGE			
		COL. B	COL. C	COL. D
		IV THERE	(V IOFO)	(Y 105-D)

		COL, B	COL. C	COL. D
		(X 10E8)	(X 10E0)	(X 10E-8)
	MEAN	4.72	11.88	2.17
	RANGE	6.93	25.00	2.37
	MAX	8.22	27.00	3.45
<b>₹</b>	MIN	1.28	2.00	1.08
NO OUTLIERS	• •	<b>–</b> .	<del>-</del>	44,55

CSCX CSC85F 21 NOV 72 17:19:26 USER CFU007 200

CARDS IN 236 OUT 0 LINES 64 PROCESSING TIME 6. 3 SECONDS

#### TEST I

COMPOUND: FDA 71-20 ORGANISM: SALMONELLA G-46

DOSE LEVEL: LD5 - 150 MG/KG

TREATMENT: IN VIVO, ORAL, SUBACUTE DATE STARTED: FEBRUARY 11, 1972

	A	В	C <sub>.</sub>	Đ
			TOTAL NO.	MUTATION
ANIMAL	RAW CFU X	TOTAL CFU X	MUTANTS X	FRE (C/B)
NUMBER	10E7/0.6ML	10E8/1.0ML	10E0/1.0ML	X 10E-8
1	49.10	8.18	5.00	•61
2	57•80	9.63	4.00	•42
3	40.40	6.73	4.00	.59
4 r	32.10	5.35	4.00	.75
2 3 4 5	33.80	5.63	8.00	1.42
6	40.60	6.77	4.00	.59
7	64.60	10.77	14.00	1.30
	IMALS EQUALS	7		
	AD ANIMALS EQUA		•	
TOTAL CEU	OUT OF RANGE	QUALS 1		
		COL. B	COL. C	COL. D

,	MEAN RANGE MAX	COL. B (X 10E8) 7.58 5.42 10.77	COL. C (X 10E0) 6.14 10.00 14.00	COL. D (X 10E-8) .81 1.00 1.42
NO OUTLIERS	MIN	5.35	4.00	.42

CSCX CSC85F 21 NOV 72 17:19:52 USER CFU007 200 CARDS IN 232 OUT 0 LINES 64 PROCESSING TIME 5.84 SECONDS

TEST I

COMPOUND: FDA 71-20 ORGANISM: SACCHARONYCES D-3

DOSE LEVEL: NEGATIVE CONTROL - WATER

TREATMENT: IN VIVO, ORAL, ACUTE DATE STARTED: FEBRUARY 18, 197

	A	. В	c	b
ANIMAL	RAW CFU X	TOTAL CFU SCREENED X	TOTAL	RECOMB/CFU
NUMBER	10E5/1.0ML	10E5/1.0ML	RECOMBINANTS /1.0ML	SCREENED X 10E-5
1	110.00	•11	1.00	9.09
2 3	110.00	•11	1.00	9.09
3	283.00	•28	4.00	14.13
4 5 6	510.00	-51	6.00	11.76
5	<b>3</b> 52.00	•35	.00	
6	120.00	•12	.00	•00
7	384.00	•38	1.00	.00
8	110.00	•11	2.00	2.60 18.18
TOTAL		1.98	15.00	
No. of As	TMALE COLLEGE	•		

NO. OF ANIMALS EQUALS 8
TOTAL SCREENED OUT OF RANGE EQUALS 2

MEAN C/MEAN 8 = 7.58

NO OUTLIERS	MEAN RANGE MAX MIN	COL. B (X 10E5) .25 .40 .51	COL. C (X 10E0) 1.88 6.00 6.00	COL. D (X 10E-5) 8.11 18.18 18.18 .00
NO OUTLIERS	MT/A	•,11	•00	•00

CSCX CSC85F 21 NOV 72 17:14:44 USER CFU007 200

CARDS IN 236 OUT 0 LINES 69 PROCESSING TIME 6.56 SECONDS

#### TEST I

COMPOUND: FDA 71-20 ORGANISM: SACCHAROMYCES D-3

DOSE LEVEL: POSITIVE CONTROL - EMS - 350 MG /KG

TREATMENT: IN VIVO, ORAL, ACUTE DATE STARTED: FEBRUARY 18, 1972

	A	В	C.	D
ANIMAL	RAW CFU X	TOTAL CFU SCREENED X	TOTAL RECOMBINANTS	RECOMB/CFU SCREENED X
NUMBER	10E5/1.0ML	10E5/1.0ML	/1.0ML	10E-5
1 2 3	180+00	.18	18.00	100.00
2	352.00	<b>.</b> 35	15.00	42.61
3	172.00	•17	12.00	69.77
4. 5	92.00	• 0 9	4.00	43.48
5	503.00	<b>-5</b> 0	16.00	31.81
6	261.00	•26	10.00	38.31
7	80.00	•08	7.00	87.50
8	98.00	• <b>1</b> 0	9.00	91.84
9	951.00	•95	8,00	8.41
TOTAL		2.69	99.00	

NO. OF ANIMALS EQUALS 9
TOTAL SCREENED OUT OF RANGE EQUALS 1

MEAN C/MEAN B = 36.82

	MEAN RANGE MAX	COL. B (X 1025) .30 .87	COL. C (X 10E0) 11.00 14.00	COL. D (X 10E-5) 57.08 91.59
NO OUTL	MIN	.95 .08	18.00 4.00	100.00 8.41

CSCX CSC85F 21 NOV 72 17:14:56 USER CFU007 200

CARDS IN 236 OUT 0 LINES 70 PROCESSING TIME 6.11 SECONDS

TEST I

COMPOUND: FOA 71-20

ORGANISM: SACCHARONYCES D-3

DOSE LEVEL: LOW - 1.5 MG/KG

TREATMENT: IN VIVO, ORAL, ACUTE

DATE STARTED: FEBRUARY 18: 1972

ANIMAL Number	RAW CFU X 10E5/1.0ML	B TOTAL CFU SCREENED X 10E5/1.0ML	C TOTAL RECOMBINANTS /1.0ML	D RECOMB/CFU SCREENED X 10E-5
2 3 4 5 6 7	382.00 451.00 213.00 130.00 152.00 381.00	.38 .45 .21 .13 .15 .38	7.00 1.00 3.00 .00 .00 4.00 3.00	18.32 2.22 14.08 .00 .00
TOTAL NO. OF AN	TMALE FOLIAL A	1.85	18,00	21.13

NO. OF ANIMALS EQUALS 7 TOTAL SCREENED OUT OF RANGE EQUALS 3

MEAN C/MEAN B = 9.72

NO OUTLIER	MEAN RANGE MAX MIN S	COL. B (X 19E5) •26 •32 •45 •13	COL. C (X 10E0) 2.57 7.00 7.00	COL. D (X 10E-5) 9.46 21.13 21.13

CSCX CSC85F 21 NOV 72 17:15:22 USER CFU007 200

CARDS IN 236 OUT O LINES 68 PROCESSING TIME 5.95 SECONDS

#### TEST I

COMPOUND: FDA 71-20

ORGANISM: SACCHAROMYCES 8-3

DOSE LEVEL: INTERMEDIATE - 15 MG/KG

TREATMENT: IN VIVO. ORAL, ACUTE

DATE STARTED: FEBRUARY 18, 1972

ANIMAL Number	A RAW CFU X 1085/1.0ML	B TOTAL CFU SCREENED X 10E5/1.0ML	C TOTAL RECOMBINANTS /1.0ML	D RECOMB/CFU SCREENED X 10E-5	
1 2 3 4 5 6 7	361.00 200.00 234.00 111.00 180.00 603.00	.36 .20 .23 .11 .18 .60	1,00 .00 .00 2,00 .00 4.00	2.77 .00 .00 18.02 .00 6.63	*
TOTAL		2.35	7.00	1	

NO. OF ANIMALS EQUALS TOTAL SCREENED OUT OF RANGE EQUALS

MEAN CYMEAN B = 2.98

	COL. B	COL. C	COL. D
MEAN	(X 10E5)	(X 10E0)	(X 10E-5)
MEAN	<b>.</b> 34	1.00	3,92
RANGE	•55	4.00	18.02
MAX	•66	4.00	18.02
MIŅ	•11	•00	.00

## \* SUMMARY WITH OUTLIERS REMOVED

MEAN C/MEAN B = 2.23

MEAN RANGE MAX MIN	COL. B (X 10E5) .37 .48 .66	COL. C (X 10E0) .83 4.00 4.00	COL. D (X 10E-5) 1.57 6.63 6.63
•	·		

CSCX CSC85F 21 NOV 72 17:15:44 USER CFU007 500

6. 1 SECONDS

_	_	_	_
_			- 1
- 2	•	•	

COMPOUND: FDA 71-20

ORGANISM: SACCHAROMYCES D-3

DOSE LE/EL: LD5 - 150 MG/KG

TREATMENT: IN VIVO, ORAL, ACUTE

DATE STARTED: FEBRUARY 18 1972

	A	B TOTAL CFU	C	D.
ANIMAL	RAW CFU X	SCREENED X	TOTAL RECOMBINANTS	RECOMB/CFU
NUMBER	10E5/1.0ML	10E5/1.0ML	/1.0ML	SCREENED X 10E-5
1	310.00	•31	1.00	3.23
2	100.00	• <b>1</b> 0	1.00	10.00
3	261.00	+26	1.00	3.83
<del>4</del> 5	131.00	•13	1.00	7.63
	<b>303.0</b> 0	•30	3.00	9.90
6	302.00	•30	2.00	6.62
7	321.00	• 32	1,00	
8	124.00	•12	1.00	3.12 8.06
TOTAL		1.85	11.00	

NO. OF ANIMALS EQUALS TOTAL SCREENED OUT OF RANGE EQUALS

MEAN CYMEAN B = 5.94

(X 10E0)	(X 10E-5)
1+38	6.55
2-00	6.88
3-00	10.00
1+00	3.12
	1.38 2.00 3.00

CSCX CSC85F 21 NOV 72 17:16: 5 USER CFU007 200

CARDS IN 236 OUT 0 LINES 72 PROCESSING TIME 6. 3 SECONDS

TEST I

COMPOUND: FOA 71-20

ORGANISM: SACCHAROMYCES D-3

DOSE LEVEL: LOW 1.5 MG/KG

TREATMENT: IN VIVO, ORAL, SUBACUTE

DATE STARTED: FEBRUARY 18: 1972

ANIMAL NUMBER	A RAW CFU X 10E5/1-0ML	B TOTAL CFU SCREENED X 10E5/1.OML	C TOTAL RECOMBINANTS /1.0ML	D RECOMB/CFU SCREENED X 10E-S
1 2 3 4 5 6 7	144.00 183.00 113.00 104.00 192.00 513.00 161.00	.14 .18 .11 .10 .19 .51	.00 1.00 .00 .00 1.00	•00 •00 8•85 •00 •00 1•95
TOTAL		1.41	2,00	***

NO. OF ANIMALS EQUALS TOTAL SCREENED OUT OF RANGE EQUALS

MEAN C/MEAN B = 1.42

MEAN RANGE MAX	COL. B (X 10E5) .20 .41 .51	COL. C (X 10E0) .29 1.00 1.00	COL. D (X 10E-5) 1.54 8.85 8.85
MIN	•10	•80	•00

## \* SUMMARY WITH OUTLIERS REMOVED

MEAN C/MEAN B = .77

MEAN	(X 10E5)	COL. C (X 10ED)	COL. D (X 10E-5)
RANGE	•22	•17	.32
	•41	1•00	1.95
MAX	•51	1.00	1.95
Min	•10	.00	

C5CX CSC85F 21 NOV 72 17:16:59 USER CFU007 200 PARDS IN 236 OUT

0 LINES 82 PROCESSING TIME

6.21 SECONDS

TEST I

COMPOUND: FDA 71-20

ORGANISM: SACCHARCMYCES D-3

DOSE LEVEL: INTERMEDIATE - 15 MG/KG

TREATMENT: IN VIVO, GRAL, SUBACUTE DATE STARTED: FEBRUARY 18, 1972

	A	8 TOTAL CFU	C <sub>.</sub> Yotal	D DECOMB (CEM
ANIMAL	RAW CFU X	SCREENED X	RECOMBINANTS	RECOMB/CFU SCREENED X
NUMBER	10E5/1.0ML	10E5/1.0ML	/1.0ML	10E-5
1	270.00	•27	2,00	7.41
2	170+00	•27 •17	1.00	5.88
3	241.00	•24	3,00	12.45
4	433.00	•43	4.00	9.24
4. 5	97.00	-10	1.00	10.31
6 7	102.00	•10	.00	•00
7	281.00	• 28	1.00	3.56
<b>8</b> .	164.00	•16	.00	• 00
TOTAL,		1.76	12,00	

NO. OF ANIMALS EQUALS TOTAL SCREENED OUT OF RANGE EQUALS 2

MEAN C/MEAN 8 = 6.83

		COL. B	COL. C	COL. D
		(X 10E5)	(X 10E0)	(X 10E-5)
	MEAN	.22	1.50	6.11
	RANGE	.34	4.00	12,45
	MAX	.43	4.00	12,45
	MIN	.10	•00	.00
NO OUTLIERS		•	•	•

CSCX C5C85F 21 NOV 72 17:17: 9 USER CFU007 200

CARDS IN 236 OUT 0 LINES 69 PROCESSING TIME

TEST I

COMPOUND: FDA 71-20 ORGANISM: SACCHAROMYCES D-3

DOSE LEVEL: LOS - 150 MG/KG

TREATMENT: IN VIVO, ORAL, SUBACUTE DATE STARTED: FEBRUARY 18, 1972

ANIMAL NUMBER	A RAW CFU X 10E5/1.0ML	B TOTAL CFU SCREENED X 10E5/1.0ML	C TOTAL RECOMBINANTS /1.0ML	D RECOMB/CFU SCREENED X 106-5
1 2 3 4 5 6 7	203.00 112.00 520.00 123.00 133.00 434.00 251.00	.20 .11 .52 .12 .13 .43	.00 2.00 3.00 2.00 3.00 1.00	.00 .00 3.85 24.39 15.04 6.91 3.98
TOTAL	•	1.78	11.00	

NO. OF ANIMALS EQUALS TOTAL SCREENED OUT OF RANGE EQUALS

MEAN C/MEAN B = 6.19

MEAN RANGE MAX MIN	COL. B (X 10E5) .25 .41 .52	COL. C (X 10E0) 1.57 3.00 3.00	COL. D (X 10E-5) 7.74 24.39 24.39
· · · · · ·		•00	•00

## \* SUMMARY WITH OUTLIERS REMOVED

MEAN C/MEAN B = 4.84

•	COL. B	COL. C	COL. D
MEAN	(X 10E5)	(X 10E0)	(X 10E-5)
RANGE	•28	1.33	4.96
MAX	.41	3.00	15.04
MIN	•52	3.00	15.04
MAN	•11	•00	•00

CSCX CSC85F 21 NOV 72 17:17:27 USER CFU007 200

CARDS IN 236 OUT 0 LINES 82 PROCESSING TIME

## 3. Host-Mediated Assay - Test II

Compound FDA 71-20 produced no significant increases in recombinant frequencies when tested against <u>Saccharomyces</u> D3. The repeat results are similar to the original results, however, the acute intermediate showed a slight increase. The lower negative control values give a better view of the compound in that there is no question of repression of recombinance as appeared in the original.



#### Evaluation of Test II

The results from the D3 repeat of compound 71-20 are acceptable although two points need to be made:

- 1. The overall increase in recoveries leading to higher colony densities on the scored plates tends to depress the recombination frequencies."

  Recoveries in the original tests were about one half of what they were in the repeat, and that is the reason for the two distinctly different negative control frequencies. Unfortunately in this run the positive control frequencies appear inconsistent with what I have just stated; however, it is clear from other similar tests both at Bionetics and at other laboratories that positive controls are likewise repressed at high colony densities on scored plates.
- 2. The impression obtained in the original test that reduced recombination resulted from treatment with compound 71-20 is not evident in the repeat. In fact, several of the dose levels show slightly increased frequencies, and the results at the acute intermediate dose may be considered a weak positive.

The overall interpretation of the original test and of the repeat are similar but not identical. The original test indicated not only negative results but even a suppression of the spontaneous recombinational events. The repeat tests suggest negative or at best weakly positive results for one or two dose levels.

David Brusick

b. HOST-MEDIATED ASSAY SUMMARY SHEETS

TEST II

CONTRACT FDA 71-268

COMPOUND FDA 71-20

SODIUM BISULFITE



SUMMARY SHEET

OUTLIERS INCLUDED

TEST II

COMPOUND: FDA 71-20

		SALMO			SACCHARONY	'CES U-3
	TA153	30	G-40	;		
	MHF (X 10E+8)	MFT/MFC	MMF (X 10F=8)	MET/HEC	MRF (X 10E-5)	MRT/MRC
ACUTE		-				
NC	1.00		1.00		2.96	
PC	0.	o.	0.	0.	34.59	11.69
AL.	<b>C</b> .	o.	ó.	0.	5.89	1.99
AI .	o.	G.	o.	o.	12.38	4.18
LDS	o.	ō.	D.	D.	3.73	1.2€
SUBACUTE						
NC	1.00		1.00		2.65	
5L	0.	٠.0	0.	0.	6.65	2.51
S 1	0.	0.	ů.	ō.	6.18	2.33
SLD5	Ď.	D.	ō.	ŏ.	9.41	3.55
PC*	o.	0.	0.	ō.	30.22	3.55 10.75
IN VITRO	TÁ1530	G-46		0-3		
		J	<b>♦</b> CONC	1 SURVIVAL	R X 10	<b>E</b> 5
NC						•

NC PC

STOP SRU'S:.6

<sup>\*</sup> Positive control performed by acute method done with subacute studies.

## SUMMARY SHEET

## OUTLIERS REMOVED.

TEST II

COMPOUND: FDA 71-20

•	TA153	SALMO: ₹n		<del>.</del>	SACCHAROMY	CES D-3
		,,	G-46			
	MMF (X 10E-8)	MFT/MFC	MMF (X 10E-8)	MFT/MFC	MRF (X 10E-5)	MRT/MRC
ACUTE						
NC PC AL Al LD5	1.00 0. 0. 0.	0. 0. 0.	1.00 0. 0. 0.	o. o. D.	2.96 32.59 5.29 12.38	11.01 1.79 4.18
E de la Compa		- •	•	0.	3.73	1.26
SUBACUTE NC SL SI SLD5 PC*	1.00 0. 0. 0.	0. 0. 0.	1.00 0. 0. 0.	0. 0. 0. 0.	2.81 6.65 6.18 8.07 30.22	2.37 2.20 2.67 10.75
IN VITRO	TA1530	G-46	<b>₹</b> CONC	0-3 § SURVIVAL	R X 10E	
NC .			-		102	-

NC PC

· P

STOP SRU'S:.6

<sup>\*</sup> Positive control performed by acute method done with subscute studies.

#### SUMMARY SHEET

OUTLIERS REBEDEED

TEST II

COMPOUND: FDA 71-20

	,	SALMO	NELLA		SACCHARONY	(CES D-3
	TA1530		6-46	<b>;</b>	**************************************	
	MMF (X 10E-8)	MFT/MFC	MMF (× 10E-8)	MFT/MFC	MRF (X 108-5)	MRT/MRC
ACUTE	•					
NC PC	1.00 0.	0.	1.00	0.	2.96 32.59	11.01
AL AI	0. 0.	0. 0.	0.	C.	5.29	1.79
LD5	ŏ.	0.	0. 0.	0. 0.	12.38 3.73	4.18 1.26
SUBACUTE						
NC	1.00	· · · · · · · · · · · · · · · · · · ·	1.00		2.81	
SL 51	0. G.	0. 0.	o.	0.	6.65	2.37
\$1.05	ŏ.	ŏ:	0.	0. 0.	6.18 8.07	2.20
IN VITRO	TA1530	 <b>c-4</b> 6		D-3		
			CONC	SURVIVAL	R X 10E	5
NC PC						

#### SUMMARY SHEET

#### OUTLIERS INCLUDED

TEST II

SALMONELLA

COMPOUND: FDA 71-20

	TA1530		G46	G-46			
_	)mif (x 10E-8)	MFT/MFC	MMF (X 10E-8)	MPT/MPC	MRF (X 10E-5)	MRT/MRC	
ACUTE						٠.	
NC ·	1.00		1.00		2.96		
PC	0.	0.	0.	0.	34.59	11.69	
AL	0.	ō.	ō.	Ġ.	5.89	1.99	
AI .	ō.	Õ,	ō.		12.38	4.18	
LD5	ů.	ō.	Ŏ.	ō.	3.73	1.26	
SUBACUTE							
NC	1,00		1.60		2,65		
SL	0.	0.	0.	0.	6.65	2.51	
SI	o.	O.	Ŏ.	o.	6.18	2.51 2.33	
SLD5	ō.	0.	ō.	ō.	9.41	3.55	
IN VITRO	TA1530	c-46	-	D~3			
		- · ·	& CONC	& CIRVIVA		r K	

SACCHAROMYCES D-3

NC

PC

STOP SRU'S:.6

COMPARASION OF

## REPEATS TO ORIGINALS

## COMPOUND FDA 71-20

## OUTLIERS INCLUDED

<u>Saccharomyces</u> D-3 Original		<u>Saccharomyces</u> D-3 Repeat		
	MMF (X 10E-8)	MFT/MFC	MMF (x 10E-8)	MFT/MFC
ACUTE		•		
NC -	7.58		2.96	
PC	36.82	4.86	34 . 59	11.69
AL	9.72	1.28	5.89	1.99
Aī	2.98	.39	12.38	4,18
HA	5,94	.78	3.73	1.26
SUBACUTE		·		
NC	7.58		2.65	
\$L	1.42	.19	6.65	2,51
51	6.83	.90	6.18	2.33
· SH	6.19	.82	9,41	3.55
PC*	-	-	30.22	10.75

<sup>\*</sup> Positive control performed by acute method done with subacute studies.

COMPARASION OF

### REPEATS TO ORIGINALS

## COMPOUND FDA 71-20

## OUTLIERS REMOVED

	Saccharomyces D-3 Original		Sacchard D-3 Re	omyces epeat
ACUTE	<del>操作</del> (X 10E-8)	MFT/MFC	7MMF (3-30f X)	MFT/MFC
NC PC	7.58 36.82	4.86	2.96 32.59	11.01
AL AI	9.72 2.23	1.28	5.29 12.38	1.79
. Ан	5.94	.78	3.73	1.26
SUBACTTE NC	7.58		2.81	
21 2F	.77 6.83	.10 .90	6.65 6.18	2.37 2.20
SH PC*	4.84	. 64	8.07 30,22	2.87 10.75

<sup>\*</sup> Positive control performed by acute method done with subacute studies.

COMPARASION OF

### REPEATS TO ORIGINALS

## COMPOUND FDA 71-20

## OUTLIERS REMOVED

	Saccharomyces D-3 Original		<u>Saccharomyces</u> D-3 Repeat	
ACUTE	MMF (X 10E-8)	MFT/MFC	MMF (X 10E-8)	MFT/MFC
NC .	7.58		2.96	
PC	36.82	4.86	32.59	11.01
AL	9.72	1.28	5.29	1.79
AI .	2.23	.29	12,38	4.18
AH	5.94	.78	3.73	1.26
SUBACUTE	<u>:</u>			
NC	7.58		2.81	
SL	.77	.10	6.65	2.37
SI	6.83	.90	6.18	2.20
SH	4.84	. 64	8.07	2.87

COMPARASION OF

## RÉPEATS TO ORIGINALS

## COMPOUND FDA 71-20

## OUTLIERS INCLUDED

	Saccharomyces D-3 Original		<u>Saccharomyces</u> D-3 Repeat		
	ACUTE	MMF (X 10E-8)	MFT/MFC	MMF (X 70E-8)	MFT/MFC
	ACUTE NC PC AL AI AH	7.58 36.82 9.72 2.98 5.94	4.86 1.28 .39 .78	2.96 34.59 5.89 12.38 3.73	11.69 1.99 4.18 1.26
•	SUBACUTE NC SL SI SH	7.58 1.42 6.83 6.19	.19 .90 .82	2.65 6.65 6.18 9.41	2.51 2.33 3.55

C. HOST-MEDIATED ASSAY DATA SHEETS

TEST II

CONTRACT FDA 71-268

COMPOUND FDA 71-20

SODIUM BISULFITE



#### TEST II

COMPOUND: FDA 71-20

ORGANISM: SACCHAROMYCES D-3

DOSE LEVEL: NEGATIVE CONTROL - SALINE (ACUTE)

TREATMENT: IN VIVO, ORAL, ACUTE DATE STARTED: FEBRUARY 12, 1973

ANIHAL NUMBER	A RAW CPU X 10E5/1.0ML	B TOTAL CFU SCREENED X 10E5/1.0ML	C TOTAL RECOMBINANTS /1.0HL	D RECOMB/CFU SCREENED X 108-5
1 2 3 4 5 6 7	418.00 687.00 984.00 317.00 328.00 309.00 340.00	.42 .69 .98 .32 .33 .31	0. 2.00 2.00 1.00 2.00 2.00	0. 2.91 2.03 3.15 6.10 6.47
TOTAL		3.38	10.00	2.94
TOTAL SCRE	IMALS EQUALS Eened out of Ra Nn B # 2	7 INGE EQUALS	3	
HO OUTLIER	MEAN RANGE MAX NIN	COL. B (X 10E5) .48 .67 .98	COL. C (X 10E0) 1.43 2.00 2.00	COL. D (X 10E-5) 3.37 6.47 6.47

SWITCH INSTRAS

TEST II

COMPOUNDS FOR 71-20

OHGANISMI SACCHARONYCES Das

DOSE LEVEL: POSITIVE CONTROL -EMS - 350 NG/AG I.M. (ACUTE)

TREATMENT: IN VIVO. ORAL, ACUTE

DATE STARTED! FEBRUARY 12, 1975

	À	TOTAL CFU	€ TATAL	RECOMS/CFU
ANIMAL	HAW CPU X	SCHEENED X	RECOMBINANTS	
HUMBER	10E5/1.0ML	10E5/1.0ML	VI-ONL	SCREENED X 106-5
4	533.00	-53	19.00	35.65
<u>£</u>	<b>317.</b> 99	.32	21.06	66.23
3	985.40	.99	34.00	34.49
4	612.00	.61	17.00	
5	767.00	.77		27.78
b	847.00		25.00	29.99
ÿ		<b>.</b> 95	25.00	<b>33.</b> 06
-	420.00	.42	10.00	47.86
¢	&37+00	• 44	24.00	23.67
TOTAL		5.32	184,00	

NO. OF ANIMALS EQUALS B TOTAL SCHEENED OUT OF HANGE EGUALS 2

MEAN CYMEAN B =

34.59

	(X 10,5)	(X 1000)	COL. U (x 105-5)
MEAN	• 64g	23.00	37.34
Rangl Hax	•67 •00	17.00	<b>⋽₿</b> •47
MIN	•99 • <b>3</b> 2	34.90 17.00	56+25
	, , , , ,		27.78

## SUMMARY WITH OUTLIERS REMOVED

HEAN CAMEAN B =

32.59

	COL.	COL. C	COL. D
346. Aug.	(X 1065)	(X 10EU)	(x 10E-51
MLAN	• <u>* * }                                   </u>	23.29	33.21
rangi. Max	• 57	17,00	15.08
MIN	<b>.99</b>	34 - 40	42.56
to Tibil	4 M 24	17.00	27.78

### TEST II

COMPOUND: FDA 71-26

ORGANISMI SACCHAROMYCES D-1

LOSE LEVEL: LOW - 148 MO/KG

TREATMENT: IN VIVO. ORAL, ACUTE

DATE STARTED! FEBRUARY 12, 1473

animal Number	RAW CFU X 1085/1.04L	TOTAL CFU SCREENED X 10ES/1.0ML	TOTAL RECOMBINANTS /1.0%L	RECOMBICEU SCREENED X 198-5
i	460.03	•48	2.00	. were
	374.00	47		4.17
7		•37	2.0y	5,39
% .5 4	493.05	.49	<b>Q</b> (i	K. 77
*	#56.00	• ពិម	4.00	8.67
ž.	<b>578.</b> 00	•58	4.00	
6	574.00	.57		5.92
7			4.04	6.97
	617.00	.62	i≩,6₩	3,24
<b>5</b>	640.03	*61	# <b>4</b> €	7.55
TATAL		4.58	27.60	

HO. OF ANTHALS EQUALS & TOTAL SCREENED OUT OF RANGE EQUALS 2

MEAN CANEAN B =

3 - 43

HEAN	(ol. 6 (x 1065) (57	CUL. C (* 1060) 3.38	COL. # (x 100+6) 5.90
HANGE	•46 •86	4.90	6.59
HAX		D. 00	9.44
MIN	.37	2,00	3.24

## \* SUMMARY WITH OUTLIERS REMOVED

MEAN CYHEAN B =

5-29

MEAN	(X 10±5)	(X 10E0)	COL- 0 (X 10E-5)
RANGE	. 57	2.00	5.34
	• 46	2.00	3.73
MAX	. 60	4.00	6.97
	. 37	2.00	3.34

56

TEST II

COMPOUND: FUA 71-20

ORGANISMI SACCHAROMYCES D-S

DOSE LEVEL: INTERNEDIATE - 15.0 MG/KG

TREATMENTS IN VIVO. CHAL. ACUTE

DATE STARTED: FEBRUARY 12. 1973

	K	ð Total öfu	TOTAL	D RECOMB/CFU
animal Humuer	HAW CFU X 10E5/1.UML	SCREENED A 10ES/1.OML	RECOMMINANTS /1.OML	SCREEMED X
į.	466.00	_49	7.00	10.40
Z	357.00	. 16	4.03	11.20
3	710.00	.71	5.00	7.04
4	357.60	• 59	4.00	20.67
5	554.00	455	9.00	16.25
Ü	962.96	• 90	6.00	8.32
7	341.09	.34	C. DU	17.60
TOTAL		3.40	47.00	

NO. OF ANIMALS EQUALS T NO. OF CONTARINATED EQUALS 1 TOTAL SCREENED OUT OF RANGE EQUALS 2

MEAN CHIEAR B =

12.30

	COL.	COL. C	coll. b
	(X LULU)	(x 1020)	(x 100-5)
MEAN	• <del>š</del> \$ 4	6.71	13.64
RANGE	, å∠	5.00	13.43
MAX	•96	9.00	20.67
MIN	•34	4.00	7.04

NO CUTLIERS

4ctaP

TEST II

COMPOUND: FOA 71-20

ORGANISMI SACCHAROMYCES D-1

COSE LEVEL! LOS - 150.0 MG/KG

TREATMENT: IN VIVO. GRAL, ACUTE

DATE STARTED: FEBRUARY 12, 1973

ANTHAL	RAW CFU X	B Yotal cfu Screeneu a	Ç Total	RECONS/CFU
NUMBER	1005/1.CML	10E5/1.UML	RECOMBINANTS /1.CML	SCREENED Y
į.	580.00	.56	4.00	6.90
ji. 3	884.93	• 45	1.00	1.t3
	487.80	440	3.00	6.15
4	707.40	.71	1.00	1.01
5	<b>910.</b> 03	.71 .91	2.00	2.20
<u>.</u>	963.40	• 96	₽.00	5.23
7	826.00	دَيْ.	2.00	3.65
TOTAL		5.36	29.00	

NO. OF ANIMALS EQUALS 7 TOTAL SCHEENED OUT OF RANGE EQUALS 3

MEAN C/MEAN B =

1.75

		COL.	CUL. C	0 - 100 (2-301 a)
	MEAN	.77	2.86	3.95
	RANGE.	a 4-51	5,00	5.77
	MAX	• ਮੌਨ	6.00	6.93
HO OUTLIERS	MIN	<b>e</b> <sup>14 j</sup> 9	1.0 <del>0</del>	1.13

. iv B

	<b>-</b> "	TEST EI	· • · · • • · · • • • • • • • • • • • •	
COMPOUND:	ŁĎY 1Í-Š0		ORGANISH! SAC	CHARONYCES D-3
DOSE LEVE	L: WEGATIVE CO	NTROL - SALIN	ε	ź,
<u> T</u> REA JMENT	IN VIVO, DRA	L. ACUTE	DATE STARTEDE	MARCH 16+ 1973
ANIMAL	A RAN ÇEU X	B Total cfu Screened X	C Totál Recombinants	D RECONS/CFU SCREENED K
NUMBER	10E5/1.0ML	IDES/1.OML	/1.0ML	ĮĢĚ÷Š
1 2 3 4 5 6 7 8	806.00 546.00 858.00 678.00 698.00 829.00	.61 .55 .56 .56 .70 .63	2.00 2.00 2.00 2.00 2.00 3.00	2.48 1.63 2.33 3.55 2.95 2.07 3.62 0. **
TOTAL		5,29	14.00	
HO. OF DEA	NALS EQUALS D ANIHALS EQU TAHINATED EQU N B =	8 ALS 1 ALS 1 2.65	· •-	
,	MEAN RANGE MAX MIN	COL. 8 (% 1965) .66 .54 .84 .31	COL. C (X 16E0) 1.75 3.00 3.00	COL. D (x 105-5) 2.45 3.62 3.62 4.
	4	SUMMARY WITH	OŅĪLIĒRS RĒMOVĒD	
MEAN CYMEN	N B =	2.81		
	Hean Range Max Min	COL. 8 (\$ 1065) -71 -31 -96	1.56 2.66 2.66 3.66 3.66	(X 10E-5) 2.80 1.79 3.62 1.83

		TEST II		•
COMPOUND	: FOA 71-20		ORGANISM: SAC	CHAROMYÇES D-
DOSE LEV	EL: POSITIVE CO	NTROL - EMS -	350 MG/KG IN (S	A+E04)
TREATHEN	T: IN VIVO, ORA	NL+ ACUTE	OATE STARTED:	MARÇH 16, 19
	ř A	8	s C	D <sup>‡</sup>
animal Humber	RAN CFU X	TOTAL CFU SCREENED X 1065/1.0ML	TOTAL RECOMBINANTS 71.0ML	RECOMB/CFU SCREENED X 106-5
1 2	921.00 1784.00	.92 1.78	13.00 39.00	14.12
3. 4.	"984.00 594.00	.98 .59	46.00 22.00	21,86 46,75 37,04
	793.00 <b>442.</b> 00	•79 •84	32.00 17.00	40.35 20.19
¥ \$	972.00 854.00	.97 .85	37.00 28.00	38.07
TOTAL		7.74	234-00	
NO. OF CO	HALS EQUALS MTAMINATED EQU EENED OUT OF R	B ALS I ANGE EUVALS	1	
***				,

		COL. B (X 10€5)	COL. C (X 10E0)	COL. D (X 10E-5)
	MEAN RANGE	97	29.25	31.39
	MAX	1.19 1.78	33.00 46.00	32.63 46.75
 M GUTI TERS	MIN	-59	13.00	14.12

TEST II

		TEST II		
COMPOUND	FDA 71-20		DRGANISM: 5AC	CHARONYCES D-3
DOSE LEVI	EL# LOW - 15 MG	∖kë		
TREATHEN	T: IN VIVO+ ORA	L. SUBAÇUTE	DATE STARTED!	MARCH 16+ 1973
ANIHAL	A RAW CFU X 10E5/1.0HL	B TOTAL CFU SCREENED X 1865/1.0ML	C TOTAL RECOMBINANTS 71.0ML	D HECOMB/CFU SCREENED X 102-5
1 2 3 4 5 6 7 8	533.00 867.00 576.00 876.00 1516.00 870.00 810.00 784.00	.53 .81 .56 .87 1.52 .87 .81	2.00 2.00 7.00 8.00 9.00 4.00 7.00	3.75 2.48 12.15 9.15 5.94 4.60 8.64 7.65
TOTAL		6-77	45.00	
NO. OF CO	NIMALS EQUALS DETAMINATED EQU REENED OUT OF R	ē•ēp Viē Ēdāvra Vra J	1	
		(X jõE2) COL• 8	COL. C	(X 10E-5)

		COL. B	COL. C	COL. D
		(Å 10E5)	(Ř 10EÖ)	(X 10E-5)
	MEAN	. 45	5,63	6.80
•	RANGE	.9¢	7,00	9.67
-	MAX	1.52	9.Q0	12.15
••	MIN	.53	2.00	2.46
B BUTI TERS			·	- · · -

### TEST II

COMPOUND!	FDA	71-20
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ORGANISM: SACCHAROMYCES D-3

DOSE LEVELI INTERMEDIATE - 15.0 MG/KG

TREATMENT: IN VIVE. ORAL. SUBACUTE DATE STARTED: MARCH 16, 1979

		_		,
	<b>A</b>	8	C C	D
		TOTAL CFU	TOTAL	RECOMB/CFU
ANIMAL	RAW CFU X	SCRÉENED X	RECOMBINANTS	SCREENED X
MUMBER	10E5/1.0ML	106571.0HL	71.OML	ÌÒĒ-5
1	675.00	<b>.67</b>	2.00	2.96
.ž	324.00	,67 ,32	0.	5.
.3	360.00	.38	2.00	5.26
.4.	354.00	.35	3.00	8.47
Ś	\$19.00	.ŚŻ	5.00	9.63
	344.60	.38	5.00	13.02
7	764.60	•76	4.00	5.26
TOTAL		3.40	21.00	
	NIMALS EQUALS	7		•
TOTAL SCI	REENED OUT OF R	YNGĒ ĒBŪVTZ	3	
WZ 444 O M45				

		COL. B (Å 10E5)	COL. C (% 10E0)	CDL. D (X 10£-5)
2	MEAN	49	3.00	6.37
<del>-</del>	Man <del>g</del> e Max	.76	5.00	13.02
	MIN	.32	5. <u>0</u> 0	13.02
NO OUTLIERS				

	FDA 71-20	TEST II	NUGAUTCH: ELF	PULBANUPER A A
			hidautau svi	CHAROMYCES D-3
DOZE TEX	EL: LD5 - 150.0	MĞ∖Kġ '		
TREATHEN	E: IN VIVO. DRA	r. zňbýčníř	DATE STARTED	MARCH 16. 1973
	Á		c	D
ANINAL	RAW CFU X	TOTAL CFU Screened X	TOTAL Recombinants	
NUMBER.	10ES/1.OML	1065/1.0HL	VI-ONF	SCREENED X
1	974.00	.97	8.00	8.21
:2 -5 -4	724.00	•72	0,00	11.05
' <u>3</u>	482.00 <b>526.</b> 00	.4ê	3.00	6.22
5	1208.00	.53 1.21	6.00	11.41
	70.00		5.00 9.00	4.14 9.81
Ţ	485.00	-48	11-00	\$\$.68 *
TOTAL		5,32	50.00	
MU. UP AN	WIMALS EQUALS	7		
	EENED OUT OF R	ANGE EQUALS	3	
	EENED OUT OF R	9.41 COL. B	COL. C	COL. D
	EENED OUT OF R	9.41 COL. 8 (X 10E5)	COL. C (X 10E0)	(X 10E-5)
	EENED OUT OF R	9.41 COL. 8 (X 10E5)	COL. C (X 10E0) 7.14	(X <sup>*</sup> 10E=5) 10.50
	MEAN RANGE	COL. 8 (X 10E5) .76 .73	COL. C (X 10EÖ) 7.14 8.00	(X <sup>*</sup> 10E-5) 10.50 18.54
	EENED OUT OF R	9.41 COL. 8 (X 10E5)	COL. C (X 10E0) 7.14	(X <sup>*</sup> 10E=5) 10.50
	MEAN RANGE MAX	COL. 8 (X 10E5) .76 .73 1.21 .48	COL. C (X 10E0) 7.14 8.00 11.00	(X 10E-5) 10.50 18.54 22.68 4.14
	MEAN RANGE MAX MIN	COL. 8 (X 10E5) .76 .73 1.21 .48	COL. C (X 10E0) 7.14 8.00 11.00 3.00	(X 10E-5) 10.50 18.54 22.68 4.14
MEAN C/HE	MEAN RANGE MAX MIN	COL. B (X 10E5) .76 .73 1.21 .48 SUNMARY WITH B.07	COL. C (X 10E0) 7.14 \$.00 11.00 3.00 OUTLIERS REHOVE	(X 10E-5) 10.50 18.54 22.68 4.14
MEAN C/HE	MEAN RANGE MAX MIN	COL. B (X 10E5) .76 .73 1.21 .48 SUMMARY WITH B.07 COL. B (X 10E5)	COL. C (X 10E0) 7.14 8.00 11.00 3.00 OUTLIERS REHOVE: COL. C (X 10E0)	(X 10E-5) 10.50 18.54 22.68 4-14 0 COL. 0 (X 10E-5)
MEAN C/HE	MEAN HEAN	COL. B (X 10E5) .76 .73 1.21 .48 SUMMARY WITH 6.07 COL. B (X 10E5)	COL. C (X 10E0) 7.14 8.00 11.00 2.00 OUTLIERS REHOVE: (X 10E0) 6.50	(X 10E-5) 10.50 18.54 22.68 4-14 0 (X 10E-5) 8.47
MEAN C/HE	MEAN HEAN RANGE	COL. B (X 1065) .76 .73 1.21 .48 SUMMARY WITH B.07 COL. B (X 1065) .81	COL. C (X 10E0) 7.14 8.00 11.00 2.00 OUTLIERS REHOVE: (X 10E0) 6.50 6.00	(X 10E-5) 10.50 18.54 22.68 4.14 0 COL. D (X 10E-5) 8.47 7.27
MEAN G/H	MEAN HEAN	COL. B (X 10E5) .76 .73 1.21 .48 SUMMARY WITH 6.07 COL. B (X 10E5)	COL. C (X 10E0) 7.14 8.00 11.00 2.00 OUTLIERS REHOVE: (X 10E0) 6.50	(X 10E=5) 10.50 18.54 22.68 4.14 0 COL. 0 (X 10E=5) 8.47

#### 4. Cytogenetics

#### a. <u>In vivo</u>

#### Acute study

The regative control group was within normal values as were the low, intermediate, and  $LD_5$  dosage level groups with respect to chromosome breaks. No other aberrations were observed. The positive control group exhibited severe chromosome damage as expected from the positive control substance TEM. The mitotic indices were within normal values. The mitotic indices of the 48-hour intermediate group and the 24- and 48-hour  $LD_5$  groups were slightly, but no significantly, elevated.

(2) The negative control group and the compound dosage level groups (low, intermediate, and  $LD_5$ ) contained cells with 6, 3, 6, and 3 percent breaks, respectively. These are within normal values. The mitotic indices were within normal ranges.

#### b. In vitro

The negative controls contained 1% cells with bridges. The only aberration observed in the compound dosage groups was 1% of cells with bridges at the high level. This is not significant. The positive control cells contained a great many aberrations as is expected with the positive control substance TEM.

CYTOGENETICS SUMMARY SHEETS

CONTRACT FDA 71-268

COMPOUND FDA 71-20

SODIUM BISULFITE



FDA 71-20 **ACUTE STUDY** METAPHASE SUMMARY SHEET

<u>Compound</u>	Dosage (mg/kg)	<u>Time</u> *	No. of <u>Animals</u>	No. of Cells	Mitotic Index %	% Cells with Breaks	Cells with Reunions	% Cells other Aber.**	% Cells with Aber.
Negative Control	saline	6	3	150	6	0	0	G	0
	saline	24	3	150	8	0	0	O	0
	saline	48	3	150	8	2	0	O	2
Low Level	1.5	6	5	250	6	0	0	0	0
	1.5	24	5	250	9	0	0	0	0
	1.5	48	5	250	10	0	0	0	0
Intermediate	15 15 15	6 24 48	5 5 5	250 250 250	8 8 6	4 0 2	. 0	_0 0	4 0 2
LD <sub>S</sub> Level	150	6	5	250	7	0	0	0	0
	150	24	5	250	10	0	0	0	0
	150	48	5	250	11	3	0	0	3
Positive Control (TEM)***	0.30	48	5	250	4	29	12	2(a)	36

<sup>\*</sup>Time of sacrifice after injection (hours).

\*\*Cells that have polyploidy (P), pulverization (pp), or greater than 10 aberrations (a).

\*\*\*Acute dose only one time. Sample taken at 48 hours.

FDA 71-20 SUBACUTE STUDY METAPHASE SUMMARY SHEET

<u>Compound</u>	Onsage* <u>(mg/kg</u> )	No. of Animals	No. of Cells	Mitotic Index %	% Cells with Breaks	X Calls with Reunions	% Cells other Aber.**	Cells with Aber.
Negative Control	saline	3	150	16	6	0	0	6
Low	1.5	5	250	10	3	0	0	3
Me d1 um	15	5	250	12	6	0	0	6
LD <sub>5</sub>	150	5	250	11	<b>3</b> ·	0	0	3

<sup>\*</sup>Dosage lx/day x 5 days \*\*Cells that have polyploidy (P), pulverization (pp), or greater than 10 aberrations (a).

FDA 71-20 ANAPHASE SUMMARY SHEET

Compound .	Dosage ** (mcg/ml)	Mitotic Index	No. of Cells	% Cells with Acentric Frag.	% Cells with Bridges	# Multipolar Cells	% Cells Other Aber.*	% Cells with Aber.
low Level	2	3	100	0	0	0	0	0
Medium Level	20	4	100	0	0	0	0	0
High Level	200	2	100	0	1	0	0	1
Negative Control	saline	2	100	0	1	0	0	1
Positive Control (TEM)	0.1	1	100	4	12 -	1	2 (pp)	16

<sup>\*</sup>Cells that have polyploidy (P), pulverization (pp), or greater than 10 aberrations (a). \*\*Cells harvested 24 hours after addition of the compound.

#### 5. Dominant Lethal Study

## Acute study

In general, significant differences between the negative control and experimental groups were shown in a few instances, but no strong indications of change were seen.

### b. Subacute study

Significant increases in both average preimplantation losses and average resorptions were shown in the experimental groups at weeks 2 and 4.



SUMMARY TABLES

CONTRACT FDA 71-268

COMPOUND FDA 71-20

SODIUM BISULFITE



TABLE I COMPOUND 20 STUDY ACUTE

#### PRETILITY INDEX

 ARITH DOSE	WEEK	BISTORICAL CONTROL	NEGATIVE CONTROL	DOSE LEVEL 1.500 MG/KG	DOSE LEVEL 15.000 MG/KG	DOSE LEVEL 150.000 MG/KG	POSITIVE CONTROL
	1:	43/ 60=0.72	13/20=0.65	8/20=0.40	16/20=0.80	13/20=0.65	15/20=0.75
	2	47/ 60=0.79	12/20=0.60	13/20=0.65	17/20=0.85	14/20=0.70	17/20=0.85
	3	53/ 60=0.89	14/20=0.70	14/20=0.70	13/20=0.65	17/20=0.85	16/20=0.80
	4	55/ 60=0.92	12/20=0.60	13/20±0.65 **	15/20=0.75	16/20=0.80	17/20=0.85
	5	52/ 60=0.87	15/20=0.75	16/20=0.80	17/20=0.85	14/20=0.70	17/20=0-85
	6	51/ 60=0.85	15/20=0.75	16/20=0.80	15/20=0.75	16/20=0.80	19/20=0.95
	7	52/ 60=0.87	15/20=0.75	15/20=0.75	18/20=0.90	14/20=0.70	19/20=0.95
	8	52/ 60=0.87	16/20=0.80	15/20=0.75	17/20=0.85	17/20=0.65	17/20=0.85

SYMBOLS OF FIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE REGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT BELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GROUP

ONE 1.\* = SIGNIFICANT AT P LESS THAN 0.05 TWO 1.\* = SIGNIFICANT AT P LESS THAN 0.01

<sup>\*</sup> SIGNIPICANTLY DIPPERENT FAOS CONTROL

I SIGHIPICANT LINEAR RELATIONSHIP WITH ARITH OR LOG DOSE (HEADING OF CULUMN)

TABLE II
COMPOUND 20 STUDY ACUTE

## AVERAGE MUSBER OF IMPLANTATIONS PER PRECHART FEMALE

النب في الأسم إلى أبيه أبي أبي ،

	PSE DOSE	BEEK	HISTORICAL CONTROL	BEGATIVE CONTROL	DOSE LEVEL 1.500 MG/KG	DOSE LEVEL 15.000 MG/KG	DOSE LEVEL 150.000 MG/KG	POSITITE CONTROL
		1	517/ 43=12.0	158/13=12.2	86/ 8=10.8	198/16=12.4	166/13=12.8	194/15=12.9
<b>t</b> 1	E 1	2	547/ 47=11.6	155/12=12.9	149/13=11.5	212/17=12.5	169/14=13.5 +aai	197/17=11.6
		3	624/ 53=11.8	181/14=12.9	188/14=13.4	168/13≈12₊9 €€I	216/17=12.7	215/16=13.4
5 11	S :	¢	642/ 55=11.7	135/12=11.3	169/13+13.0	192/15=12.0	213/16=13.3 el .*eel	197/17=11.6
		5	619/ 52=11.9	182/15=12.1	196/16=12.3	210/17=12.4	183/14=13.1	194/17=11.4
		6	608/ 51=11.9	179/15=11.9	178/16=11.1	188/15=12.5	198/16=12.4	228/19=12.0
		7	634/ 52=12.2	182/15=12.1	180/15*12.0	226/18=12.6	181/14=12.9	223/19=11.7
		8	605/ 52=11.6	197/16±12.3	182/15=12.1	209/17=12.3	208/17×12.2	196/17±11.5

SYMBOLS ON PIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIPPERENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GROUP

8 AND \* = INO-PAILED TEST 1 AND a = ONE-TAILED TEST

ONE 1,  $\epsilon$ ,  $\epsilon$ ,  $\star$  = SIGNIPICANT AT P LESS THAN 0.05 TWO 1,  $\epsilon$ ,  $\epsilon$ ,  $\star$  = SIGNIPICANT AT P LESS THAN 0.01

\*. SIGNIPICANTLY DIFFERENT PROM CONTROL E.! SIGNIPICANT RELATIONSHIP WITH ARITH OR LOG DOSE (HEADING OF COLUMN)

# TABLE III COMPOUND 20 STUDY ACUTE

## AVERAGE CORPORA LUTEA PER PREGNANT FREALS

LOG DOSE	ARITE DOSE	BER	HISTORICAL CONTROL	CONTROL BEGITI VR	DOSE LEVEL 1.500 HG/KG	DOSE LEVEL 15.000 MG/KG	150.000 MG/KG	POSITIVE
: 1		12	546/ 43=12.7	188/13=14.5 +aI	108/.8=13.5	225/16=14.1	182/13=14.0 ƏI	204/15=13.6
1134	1133	2	593/ 47=12.6	175/12=14.6	172/13=13.2	229/17=13.5	209/14±14.9 **a	237/17=13.9 ai
		3	673/ 53=12.7	188/14=13.4	206/14=14.7	186/13=14.3 aai a	237/17±13.9	224/16=14.0 *aI
\$611	£ 1	4	689/ 55=12.5	160/12=13.3	180/13=13.9	205/15=13.7	229/16=14.3 ***	203/17=11.9 ai
,		5	666/ 52=12.8	190/15-12.7	200/16=12-5	210/17=12.4	186/14=13.3	203/17=11.9
1		6	647/ 51=12.7	179/15=11.9	186/16=11.6 aD	194/15=12.9 <del>3</del>	1 204/16=12.8	229/19=12.1
A. (1) (A. 187)		7	664/ 52=12.8	188/15=12.5	182/15=12.1	233/18=12.9	167/14=13.4	225/19=11.8 aD
i  -  -  -		8	660/ 52=12.7	202/16=12.6	193/15=12.9	215/17=12.7	210/17=12.4	202/17=11.9

SYMBOLS ON PIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GROUP

S AND \* = TWO-PAILED TEST I AND a = ONE-TAILED TEST

ONE 1.6.4. \* SIGNIFICABLE AT P LESS THAN 0.05 TWO 1.6.4. \* = SIGNIFICABLE AT P LESS THAN 0.01

<sup>\*, &</sup>amp; SIGNIPICABLY DIFFERENT PROM CONTROL 6,1 SIGNIPICABL ARLATIONSHIP WITH ARITH OR LOG DOSE (HEADING OF COLUMN)

TABLE IV
COMPOUND 20 STUDY ACCTR

#### AVERAGE PREIMPLANTATION LOSSES PER PREGNANT FREALE

DOSE DO	eite Se	UBEK .	mistorical Control	COSTROL	1.500 MC/KC	15.000 BG/KG	150.000 MG/RG	COMINOT
		1.	29/ 43= 0.7	30/13= 2.3 *aI	22/ 8= 2.8 **a	27/16= 1.7 aI	16/13= 1.2	10/15= 0.7ap
		2	46/ 47= 1.0	20/12= 1.7	23/13= 1.8	17/17= 1.0	20/14= 1-9	40/17= 2.4
		3	49/ 53= 0.9	7/14= 0.5	18/14= 1.3	18/13= 1.4* <b>3</b> I	21/17= 1.201	9/16= 0.6
		4	47/ 55= 0.9	25/12= 2.1	11/13= 0.9	13/15= 0.9	16/16= 1.0	6/17= 0.4+aD
<b>56</b> 11		5	47/52= 0.9	8/15= 0.5	4/16= 0.3 *ap	0/17= 0.0 **a	3/14= 0.2 Pap *881	9/17= 0.5 0
		6	39/ 51= 0.8	0/15= 0.0 **a	8/16= 0.50I	6/15= 0.4	6/16= 0.4	1/19= 0.1 **#i
		7	30/ 52= 0.6	6/15= 0.4	2/15= 0.1 +ap	7/18= 0.4	6/14= 0.4	2/19= 0.1ab **a;
3 1133	1	8	55/ 52= 1.1	5/16= 0.3 *aai	11/15= 0.7	6/17= 0.4 *an	2/17= 0.1. ++a	

SYMBOLS ON PIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIPPERENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS OF SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GHOUP

& AND \* = TWO-TAILED TEST
! AND \* = ONE-TAILED TEST

ONE 1.6.2.\* = SIGNIFICANT AT P LESS THAN 0.05 TWO 1.6.2.\* = SIGNIFICANT AT P LESS THAN 0.01

<sup>\*.</sup> a SIGNIPICANTLY DIFFERENT FROM CONTROL

<sup>6.1</sup> SIGNIFICANT RELATIONSHIP WITH ARITH OR LOG DOSE (BEADING OF COLUMN)

TABLE V
COMPOUND 20 STUDY ACUTE

# AVERAGE RESORPTIONS (DEAD IMPLANTS) - PER PREGNANT PREALE

OG AR		MISTORICAL CONTROL	COLLEGE	DOSE LEVEL 1.500 MG/KG	DOSE LEVEL 15.000 MG/KG	DOSE LEVEL 150.000 HC/KG	POSITIVE CONTROL
. I	1:	9/:43=0,21	4/13=0.31	1/ 8=0.13	11/16=0.69 ƏI	8/13=0.62	3/15=0.20
	2	20/ 47=0.43	7/12=0.59	8/13=0.62	14/17=0.63	12/14=0.86	26/17=1.53*aáI **aaI
	3	25/.53=0.48	17/14=1.22 **aa	13/14=0.93 I <b>3</b> I	10/13=0.77	12/17=0.71	23/16=1.44 *aI
	4	27/ 55=0.50	4/12=0.34	18/13=1.39*aal *aal	13/15=0.8701	8/16±0.50	50/17=2.95**@@I **@@I
	5	28/ 52=0.54	10/15= <b>0.</b> 67	13/16=0.82	8/17=0.48	4/14=0.29	30/17=1.77*aI **aaI
	6	27/ 51=0.53	7/15=0-47	5/16=0.32	2/15=0.14 **aa	8/16=0.50 D	4/19=0.22 ≠ep
	7	32/ 52=0.62	2/15≈0.14 **aa	2/15=0.14 D **aa	5/18=0.28 D #D	4/14=0.29	2/19≖0.11. **aab
	8	30/ 52=0.58	8/16=0.50	6/15=0.40	7/17=0.42	11/17=0.65	13/17=0.77

SINBOLS ON PIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIPPERENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT BELATIONSHIPS AND DIPPERENCES USING THE HISTORICAL CONTROL GROUP

& AND \* = TWO-TAILED TEST ! AND & = ONE-TAILED TEST

ONE 1.6.0.\* = SIGNIFICANT AT P LESS THAN 0.05
TWO 1.6.0.\* = SIGNIFICANT AT P LESS THAN 0.01

<sup>\* .</sup> a SIGNIFICANTLY DIFFERENT PRON CONTROL

E.! SIGNIPICANT RELATIONSHIP WITH ARITH OR LOG DOSE (HEADING OF COLUMN)

COMPOUND 20 STUDY ACUTE

#### PROPORTION OF PRHALES WITH OWN OR HORE DEAD IMPLANTATIONS

LOG DOSE	APITH Dose	TREK	ELSTORICAL CONTROL	DEGATIVE CONTROL	DOSE LEVEL 1.500 EG/KG	DOSE LEVEL 15.000 EG/EG	DOSE LEVEL 150.000 BG/EG	POSITIVE CONTROL
		1.	9/ 43=0.21.	4/13=0.31	1/ 8=0.13	7/16=0.44	5/13=0.39	3/15=0.20
		Ž	14/ 47=0.30	5/12=0-42	6/13=0.47	8/17=0.48	6/14=0.43	14/17=0.83*
		3	16/ 53=0-31	10/14=0.72	8/14=0.58	5/13=0.39	7/17=0.42	9/16=0.57
	-	4	21/ 55=0.39	4/12=0.34	9/13=0.70	9/15=0.60	6/16=0.38	16/17=0.95** **
	-	5	18/ 52=0.35	6/15=0.40	8/16=0.50	7/17=0.42	3/14=0.22	13/17=0.77*
		6	21/.51=0.42	5/15=0.34	4/16=0.25	1/15=0.07	5/16=0.32	4/19=0.22
		7.	22/ 52=0.43	2/15=0.14 *	1/15=0.07	4/18=0.23	3/14=0.22	1/19=0.06
		8	20/ 52=0.39	6/16=0.38	6/15=0.40	6/17=0.36	7/17=0.42	8/17=0.48

SIMBOLS ON PIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE REGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GROUP

OBE !.\* = SIGNIFICANT AT P LESS THAN 0.05 TWO !.\* # SIGNIFICANT AT P LESS THAN 0.01

<sup>\*</sup> SIGNIPICANTLY DIFFERENT FROM CONTROL

<sup>!</sup> SIGHTPICANT LIBEAR RELATIONSHIP WITH ARITH OR LOG DOSE (READING OF COLUMN)

COMPOUND 20 STUDY ACRES

# PORPORTION OF PERALES WITH TWO OR MORE DEAD IMPLANTATIONS

g Belg Se Dosi		EISTORICAL CONTROL	CONTROL	DOSE LEVEL 1.500 EG/EG	DOSE LEVEL 15.000 AG/EG	DOSE LEVEL 150.000 EG/KG	CONTROL
	1:	0/ 43=0.0	0/13=0.0	0/ 8=0.0	3/16=0 <b>.</b> 19	1/13=0.08	0/15-0-0
	2	6/ 47=0.13	2/12=0.17	2/13=0.16	3/17=0.18	3/14=0.22	9/17=0-53*: **.
	3	7/ 53=0.14	5/14=0.36	4/14=0.29	4/13=0.31	4/17=0.24	6/16=0.38
	4	6/.55=0.11	0/12=0.0	6/13=0-47** **	3/15=0.20	2/16=0.13	11/17=0.65**
	5	8/ 52=0.16	3/15=0.20	3/16=0.19	1/17=0.06	1/14=0.08	7/17=0.42
	6	6/ 51=0.12	2/15=0.14	1/16=0.07	1/15=0.07	2/16=0.13	0/19=0.0
	7.	6/ 52=0.12	0/15=0.0	1/15=0.07	1/18=0.06	1/14=0.08	1/19=0.06
	6	8/.52=0.16	1/16=0-07	0/15=0.0	1/17=0.06	3/17=0.18	2/17=0.12

SYMBOLS OF FIRST LINE DENOTE SIGNIFICANT BELATIONSHIPS AND DIFFRRENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIPPERENCES USING THE HISTORICAL CONTROL GROUP

ONE 1.\* = SIGNIFICANT AT P LESS THAN 0.05 TWO 1.\* = SIGNIFICANT AT P LESS THAN 0.01

<sup>\*</sup> SIGNIFICANTLY DIPPERENT FROM CONTROL

<sup>1</sup> SIGNIFICANT LINEAR RELATIONSHIP WITH ARITH OR LOG DOSE (HEADING OF COLUMN)

TABLE VIII
COMPOUND 20 STUDY ACUTE

#### DEAD IMPLIETS / TOTAL IMPLIETS

VEEK	MISTORICAL Control	CONTROL	DOSE LEYEL 1.500 NG/KG	DOSE LEVEL 15.000 MG/KG	DOSE LEVEL 150.000 MC/KG	POSITIVE CONTROL
1.	9/ 517=0.02	4/158=0.03	1/ 86=0.02	11/198=0.06	8/166=0.05	3/194=0.02
2	20/ 547=0.04	7/155=0.05	8/149=0.06	14/212=0.07	12/189=0.07	26/197=0.14
3	25/ 624=0.05	17/181=0.10	13/188=0.07	10/168=0.06	12/216=0.06	23/215=0.11
•	27/ 642=0.05	4/135=0.03	18/169=0.11	13/192=0.07	8/213=0.04	50/197=0.26
5	28/ 619=0.05	10/182=0.06	13/196=0.07	8/210=0.04	4/183=0.03	30/194=0.16
6	27/ 608=0.05	7/179=0.04	5/178=0.03	2/188=0.02	8/198=0.05	4/228=0.02
7	32/ 634=0.06	2/182=0.02	2/180=0.02	5/226=0.03	4/181=0 <b>.</b> 03 -	2/223=0.01
8	30/ 605±0.05	8/197=0.05	6/182=0.04	7/209=0.04	11/208=0.06	13/196=0.07

SYMBOLS OF PIRST LINE DENOTE SIGNIFICANT DIFFERENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT DIFFERENCES USING THE HISTORICAL CONTROL GROUP

<sup>• =</sup> TWO-TALLED TEST

<sup>&</sup>amp; - ONE-TAILED TEST

ONE \*. . . SIGNIFICANT AT P LESS THAN 0.05 TWO \*. . . SIGNIFICANT AT P LESS THAN 0.01

of o Significantly Different Paon Control

COMPOUND 20 STUDY SUBACUTE

#### PERTILITI INDEX

ARITH DOSE	VESK .	EISTORICAL CONTROL	CONTROL	DOSE LEVEL 1.500 mg/kg	DOSE LEVEL 15.000 EG/EG	DOSE LEVEL 150.000 MG/KG
	1.	44/.60=0.74	14/20=0.70	16/20=0.80	14/20=0.70	12/20=0.60
:	2	44/ 60=0.74	16/20=0.80	18/20=0.90	16/20=0.80	11/20=0-55
-	3	48/.60=0.80	15/20=0.75	18/20=0.90	13/20=0.65	18/20=0.90
	• .	48/ 60=0.80	15/20=0.75	17/20=0.65	18/20=0.90	14/20=0.70
	5	<b>48/</b> 60=0.80	15/20= <b>0.</b> 75	18/20=0.90	14/20=0.70	15/20=0.75
	6	50/ 60*0.84	17/20=0.85	19/20=0.95	17/20=0.85	19/20=0-95
	7	49/.58=0.85	19/20=0.95	17/20±0.85	17/20=0.85	15/20=0.75

SYMBOLS ON FIRST LINE DENOTE SIGNIPICANT RELATIONSHIPS AND DIFFERENCES USING THE REGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GROUP

ONE 1,\* = SIGNIFICANT AT P LESS THAN 0.05 TWO 1,\* = SIGNIFICANT AT P LESS THAN 0.01

<sup>\*</sup> SIGNIFICANTLY DIFFERENT FROM CONTROL

I SIGNIFICANT LINEAR RELATIONSHIP WITH ABITH OR LOG DOSE (HEADING OF COLUMN)

TABLE II
COMPOUND 20 STODY SUBACUTE

# ATREASE MUEBRE OF IMPLANTATIONS PER PRECHAM PRELIE

-	ARITS DOSE	E RRE	HISTORICAL CONTROL	PRCATITE COSTROL	DOSE LEVEL 1.500 MG/KG	DOSE LEVEL. 15.000 MG/KG	150.000 MG/KG
		1:	492/ 44=11.2	173/14=12.4	195/16=12.3	173/14=12.4 a:	150/12=12 <b>.</b> 5
		2	540/ 44-12.3	205/16=12.8	237/18=13.2	196/16=12.3	144/11=13-1 .
1	1 1	3	580/ 48=12.1	175/15=11.7	217/18=12-1	163/13=12.5	239/18=13.3 <b>a</b> I <b>*a</b> I
		4	561/ 48=11.7	198/15=13.2	202/17=11.9	221/18=12.3	179/14=12.8 ai
		5	579/ 48=12.1	185/15=12.3	226/18=12.6	178/14=12.7	171/15=11.4
		6	610/.50=12.2	215/17=12.7	241/19=12.7	224/17=13.2 3	231/19=12.2 I
1		7	545/ 49=11.1	249/19×13.1 **aa	192/17±11.33D	211/17=12.4	183/15=12.2 I ƏI

SYMBOLS ON FIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE REGATIVE CONTROL GROUP

SIMBOLS ON SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GROUP

& AND \* \* TWO-TAILED TEST ! AND & \* ONE-TAILED TEST

ONE !. E. a. \* = SIGNIFICANT AT P LESS THAN 0.05 TWO !. E. a. \* = SIGNIFICANT AT P LESS THAN 0.01

<sup>\*, &</sup>amp; SIGNIPICABILY DIPPERENT FROM CONTROL E.! SIGNIPICANT RELATIONSHIP WITH ARITH OR LOG DOSE (HEADING OF COLUMN)

TABLE III
COMPOUND 20 STUDY SUBACUTE

# AVERAGE CORPORA LUTEA PER PREGNANT FREALE

ARITE DOSE	HEEL	EISTORICAL CORTROL	DEGATIVE CONTROL	DOSE LEVEL 1.500 MG/KG	15.000 MG/KG	150.000 MG/KG
11 3	1.	523/-44=11-9		219/16=13.7	196/14=14.0 æi ••	176/12=14.7 aai <b>+aa</b> i
	2	566/ 44=12.9	221/16=13.8	281/18=15.6aI **a	220/16=13.8 ai	152/11=13.8
6 1	3	612/ 48=12.8	205/15=13.7	293/18=12.9	182/13±14.0.	258/18=14.3 *@#I
	4	594/ 48=12.4	198/15=13.2	235/17=13.8 *#I	247/18≠13.7 ∂3	188/14=13.4 ai
	5	605/ 48=12.6	196/15=13.1	233/18=12.9	185/14=13.2	186/15=12.4
	6	641/ 50=12.8	220/17=12.9	251/19=13.2	226/17=13.3	245/19=12.9
	7	583/ 49=11.9		209/17±12.3@D	218/17±12.8 *8	183/15=12.2 <b>3</b> D
	DOS E	DOSE WERE  1	DOSE WERE CONTROL  1 523/ 44=11.9  5 11  2 566/ 44=12.9  3 612/ 48=12.8  5 1  4 594/ 48=12.4  5 605/ 48=12.6  6 641/ 50=12.8	DOSE HERE CONTROL CONTROL  1 523/ 44=11.9 201/14=14.4  5 11  2 566/ 44=12.9 221/16=13.8  3 612/ 48=12.8 205/15=13.7  6 1  4 594/ 48=12.4 198/15=13.2  5 605/ 48=12.6 196/15=13.1  6 641/ 50=12.8 220/17=12.9  7 583/ 49=11.9 258/19=13.6	DOSE HERE CONTROL CONTROL 1.500 MG/KG  1 523/ 44=11.9 201/14=14.4 219/16=13.7 ***a  2 566/ 44=12.9 221/16=13.8 281/18=15.6aI ***a  3 612/ 48=12.8 205/15=13.7 233/18=12.9  6 1  4 594/ 48=12.4 198/15=13.2 235/17=13.8 **a  5 605/ 48=12.6 196/15=13.1 233/18=12.9  6 641/ 50=12.8 220/17=12.9 251/19=13.2	DOSE HERE CONTROL CONTROL 1.500 NG/NG 15.000 NG/NG  1 523/ 44=11.9 201/14=14.4 219/16=13.7 196/14=14.0 ****  2 566/ 44=12.9 221/16=13.8 281/18=15.6aI 220/16=13.8 ****  3 612/ 48=12.8 205/15=13.7 233/18=12.9 182/13=14.0 all  4 594/ 48=12.4 198/15=13.2 235/17=13.8 247/18=13.7 all  5 605/ 48=12.6 196/15=13.1 233/18=12.9 185/14=13.2  6 641/ 50=12.8 220/17=12.9 251/19=13.2 226/17=13.3  7 583/ 49=11.9 258/19=13.6 209/17=12.3aD 218/17=12.8

SYMBOLS ON PIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIPPERENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS ON SECOND LIBE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GROUP

6 AND \* = TWO-TAILED TEST 1 AED 3 = ONE-TAILED TEST

ONE 1.6.2. \* = SIGNIFICANT AT P LESS THAN 0.05 TWO 1.6.2. \* = SIGNIFICANT AT P LESS THAN 0.01

\*. B SIGNIPICANTLY DIFFERENT PAON CONTROL 5,1 SIGNIFICANT RELATIONSHIP WITH ARITH OR LOG DOSE (READING OF COLUMN)

TABLE IT COMPOUND 20 STUDY SUBACUTE

#### AVERAGE PREIMPLANTATION LOSSES PER PREGNANT PROALE

LOG DOSE	ARITE Dose	WEEK	HISTORICAL CONTROL	REGATIVE CONTROL	DOSE LEVEL 1.500 MG/KG	DOSE LEVEL 15.000 MG/KG	150.000 MG/KG
<b>8611</b>	1133	1.	31/ 44= 0.7	28/14= 2.0 *ai	24/16= 1.5	23/14= 1.6	26/12= 2.2 **aai
Y Prove	6 1	2	26/ 44= 0.6	16/16= 1.0	44/18= 2.4 <b>**</b>	_	8/11= 0.7 *aa1
		3	32/ 48= 0.7	30/15= 2.0 *aa:	16/18± 0.9 I	19/13= 1.5 9	
		4	33/ 48= 0.7	0/15= 0.0 **a	-		*301 9/14= 0.6**00I I
HERMAN CALLS		5	26/ 48= 0.5	11/15= 0.7	7/18= 0.4	7/14= 0.5	15/15= 1.0
egge-gaziş		6	31/ 50= 0.6	5/17= 0.3	10/19= 0.5	2/17= 0.1	14/19= 0.7 *āāD
1 6611	8 11	7	38/ 49= 0.8	9/19= 0.5	17/17= 1.0	7/17= 0.4 a	

SYMBOLS ON FIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE NEGATIVE CONTROL GROUP

STABOLS ON SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GROUP

6 AND \* = TWO-TAILED TEST ! AND 8 = ONE-TAILED TEST

ONE 1.6.4.\* = SIGNIPICANT AT P LESS THAN 0.05 TWO 1.6.4.\* = SIGNIPICANT AT P LESS THAN 0.01

\*. SIGNIFICARTLY DIFFERENT FROM CONTROL

8,! SIGNIFICANT ARLATIONSHIP WITH ARITH OR LOG DOSE (HEADING OF COLUMN)

COMPOSED 20: STUDY SUBJECTE

## AVERAGE EXSORPTIORS (DEAD IMPLANTS) PER PRECEAST FRUITE

	arite Dose	WEEK	HISTORICAL: CONTROL	Cominor Cominor	DOSE LEVEL 1.500 MG/KG	DOSE LEVEL 15.000 MG/KG	DOSE LEVEL 150.000 MC/KG
e 11	<b>.</b> .	11	12/.44=0.28	11/14=0.79 aI	14/16=0.88 *aaI	13/14=0.93	18/12=1-50
		2	21/.44=0.48	4/16=0.25	31/18=1.73**aaz *aI	(17/16=1.07 <b>++33</b> ) (17/16=1.07 <b>++33</b> )	
		3.	31/ 48=0.65	7/15=0.47	14/18=0.78	5/13=0.39	11/18=0.62
		4	20/ 48=0.42	4/15=0.27	17/17=1.00*aI *aI	20/18=1.12*a3I *aI	7/14=0.50
		5	34/ 48=0.71	9/15= <b>0.</b> 60	7/18=0.39	6/14=0.43	11/15=0.74
		. 6	25/ 50=0.50	11/17=0.65	8/19=0.43	6/17=0.36	11/19=0.58
		7	36/ 49±0.74	5/19=0.27	13/17=0.7731	11/17=0.65	9/15=0.60

SYMBOLS OF FIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS OF SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GROUP

& AND \* = TWO-TAILED TEST 1 AND 3 = ONE-TAILED TEST

ONE !.E.a.\* = SIGNIFICANT AT P LESS THAN 0.05 THO !.E.a.\* = SIGNIFICANT AT P LESS THAN 0.01

\*. a SIGNIFICANTLY DIFFERENT FROM CONTROL 6.1 SIGNIFICANT RELATIONSHIP WITH ABITH OR LOG DOSE (SEADING OF COLUMN)

TABLE VI COMPOUND 20 STUDY SUBACUTE

# PROPORTION OF PERALES WITH ONE OR MORE DEAD IMPLANTATIONS

G SB	aritu Dose	WEEK.	HISTORICAL CONTROL	regative Control	Dose Level 1.500 mg/kg	DOSE LEVEL 15.000 MG/KG	DOSE LEVEL 150.000 Mg/KG
		1.	12/ 44=0.28	7/14=0.50	9/16=0.57	6/14=0.43	7/12=0.59
		2	16/ 44=0.37	4/16=0.25	12/18=0.67*	12/16=0.75**	7/11=0.64*
		3	20/ 48=0.42	6/15±0.40	10/18=0.56	4/13=0.31	8/18=0.45
		4	13/ 48=0.28	3/15=0.20	10/17=0.59*	10/18=0.56*	4/14=0.29
	•	5	23/ 48=0.48	8/15=ú.54	5/18=0.28	5/14=0.36	6/15=0.40
		6	19/ 50=0.38	5/17=0.30	7/19=0.37	6/17±0.36	6/19=0.32
		7	15/ 49±0.31	5/19=0.27	9/17=0.53	7/17=0.42	7/15=0.47

SIMBOLS ON FIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE REGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GROUP

ONE  $t_*$ \* = SIGNIPICANT AT P LESS THAN 0.05 TWO  $t_*$ \* = SIGNIFICANT AT P LESS THAN 0.01

<sup>\*</sup> SIGNIFICANTLY DIPPERBNT PROM CONTROL

I SIGNIFICANT LINEAR RELATIONSHIP WITH ARITH OR LOG DOSE (HEADING OF COLUMN)

TABLE VII STUDY SUBACUTE COMPOUND 20

# PORPORTION OF FEMALES WITH TWO OR HORE DEAD IMPLANTATIONS

	ARITH DOSE	BEEK	HISTORICAL CONTROL	REGATIVE CONTROL	Dose level 1.500 mg/kg	DOSE LEVEL 15.000 MG/KG	DOSE LEVEL 150.000 Mg/Kg
		1	0/ 44=0.0	3/14=0.22	5/16=0.32 **	2/14=0.15	1/12=0.09
		2	3/.44=0.07	0/16=0.0	5/18×0.28*	5/16=0.32*	1/11=0.10
		3	7/ 48=0.15	1/15=0.07	2/16=0.12	1/13=0.08	2/18=0.12
		4	6/ 48=0.13	1/15=0.07	4/17=0.24	6/18=0.34	1/14=0.08
		5	9/ 48=0.19	1/15=0.07	2/18=0.12	1/14=0.08	2/15=0.14
	: 1	6	4/ 50=0.08	3/17=0.18	1/19=0.06	0/17=0.0	4/19=0.22
1	ı	7	10/ 49=0.21	0/19=0.0 +	3/17=0.18	4/17=0.24 <b>*</b>	1/15=0.07

SYMBOLS ON PIRST LINE DENOTE SIGNIPICANT RELATIONSHIPS AND DIPPERENCES USING THE BEGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GROUP

ONE 1. \* = SIGNIPICANT AT P LESS THAN 0.05 TWO 1,\* = SIGNIPICANT AT P LESS THAN 0.01

<sup>\*</sup> SIGNIPICANTLY DIFFERENT PROM CONTROL

<sup>!</sup> SIGNIFICANT LINEAR RELATIONSHIP WITH ARITH OR LOG DOSE (HEADING OF COLUMN)

# TIBLE VIII COMPOUND 20 STUDY SUBACUTE

# DEAD IMPLANTS / TOTAL IMPLANTS

#2BK	MISTORICAL CONTROL	PRGATIVE CONTROL	DOSE LEVEL 1.500 MG/KG	DOSE LEVEL 15.000 NG/KG	DOSE LEVEL 150.000 MG/KG	
1.	12/ 492=0.03	11/173=0.07	14/195=0.08	13/173=0.08	18/150-0-12	
2	21/ 540=0.04	4/205=0.02	31/237=0.14	17/196=0.09	8/144=0.06	
3 -	31/.580=0.06	7/175±0.04	14/217=0.07	5/163=0.04	11/239=0.05	
4	20/ 561=0.04	4/198*0.03	17/202=0.09	20/221=0.10	7/179=0.04	
5	34/ 579=0.06	9/185=0.05	7/226=0.04	6/178=0.04	11/171=0.07	
6	25/ 610=0.05	11/215=0.06	8/241=0.04	6/224=0.03	11/231=0.05	
7	36/ 545=0.07	5/249±0.03	13/192=0.07	11/211=0.06	9/183=0.05	

SYMBOLS ON PIRST LINE DENOTE SIGNIPICANT DIPPERENCES USING THE REGATIVE CONTROL GROUP

STUBOLS ON SECOND LINE DENOTE SIGNIFICANT DIPPERENCES USING THE HISTORICAL CONTROL GROUP

ONE \*, a = SIGNIPICANT AT P LESS THAN 0.05 TWO \*, a = SIGNIPICANT AT P LESS THAN 0.01

<sup>\* =</sup> TWO-TAILED TEST a = ONE-TAILED TEST

<sup>\*,</sup> a SIGNIPICANTLY DIFFERENT PAON CONTROL

#### **APPENDICES**

#### II. MATERIALS AND METHODS

#### A. Animal Husbandry

Animals (Rats and Mice)

Ten to twelve week old rats (280 to 350 g) and male mice (25 to 30 g) were fed a commercial 4% fat diet and water ad libitum until they were put on experiment. Flow Laboratories random-bred, closed colony, Sprague-Dawley CD strain rats were used in the cytogenetic studies. Flow Laboratories ICR male mice were employed in the Host-Mediated Assay.

Preparation of Diet

A commercial 4% fat diet was fed to all animals. Periodic tests to verify the absence of coliforms, <u>Salmonella</u> and <u>Pseudomonas</u> sp. were performed.

#### 3. Husbandry

Animals were held in quarantine for 4-11 days. Mice were housed five to a cage and rats one to five to a cage. Animals were identified by ear punch. Sanitary cages and bedding were used, and changed two times per week, at which time water containers were cleaned, sanitized and filled. Once a week, cages were repositioned on racks; racks were repositioned within rooms monthly. Personnel handling animals or working within animal facilities were head coverings and face masks, as well as suitable garments. Individuals with respiratory or other overt infections were excluded from the animal facilities.

### Dosage Determination

1. Acute  $\mathrm{LD}_{50}$  and  $\mathrm{LD}_{5}$  Determination Since the compounds proposed for testing are included in



the food additive regulations as "generally recognized as safe" (GRAS), it was expected that a large number of them would be sufficiently non-toxic so that determination of a  $LD_{50}$  or a  $LD_{5}$  would be of no practical value. In fact, this has been our experience with previously tested compounds from this list. In the case of these relatively non-toxic compounds, attempts were made to assure that the amounts to be administered would not affect the animals by means (mechanical, physical, etc.) related to their bulk rather than to their toxicity. In the cases of certain compounds where a  $LD_{50}$  or a  $LD_{5}$  could not be determined, an exceedingly high concentration, 5 g/kg, was employed and accepted as the  $LD_{5}$  level. In cases where the toxicity was high enough to allow determination of a  $LD_{5}$ , the following protocol was used.

Thirty rats of the strain chosen for studies described below and of approximately the age and weight specified were assigned at random to six groups. Each group was then given, using the chosen route of administration, one of a series of dosages of the test compound following a log-arithmic dosage scheme. The series of dosages were derived from a consideration of whatever toxicity information was available for the particular test compound. The objective in selecting dosages was to choose values which would cause mortalities between 10% and 90%.

When information was inadequate to derive a suitable series of dosages, five rats were used to identify the proper range. Each of these was given one of a widely spaced (differing by 10X) series of doses. This was confidently expected to suffice for derivation of the series of dosages to be used in the  $LD_{SO}$  determination.



The mortalities observed when the series of dosages were given to the 30 rats were then subjected to a probit analysis and calculation of  $LD_{50}$ ,  $LD_{5}$ , slope and confidence limits by the method of Litchfield and Wilcoxon. The highest dose level used was either a finite  $LD_{5}$  or 5000 mg/kg. The intermediate level used was either 1/10 of the finite  $LD_{5}$  or 2500 mg/kg. The low level used was either 1/100 of the finite  $LD_{5}$  or 30 mg/kg.

#### Subacute Studies

Subacute doses were identical to those used in the acute studies. Each subacute study animal was given the acute dosage once a day for each of five consecutive days (24 hours apart).

# C. <u>Mutagenicity Testing Protocols</u>

#### Host-Mediated Assay

Flow Laboratories ICR random-bred male mice were used in this study. In the acute and subacute studies ten animals, 25-30 g each, were employed at each dose level. Solvent and positive controls were run at all times. The positive control (dimethyl nitrosamine) was run by the acute system only at a dose of 100 mg/kg for Salmonella. For yeast, ethyl methane sulfonate (EMS) intramuscularly injected at a dose of 350 mg/kg was used. The solvents used and the toxicity data are presented in the Results and Discussion Section of the report.

The indicator organisms used in this study were: (1) two histidine auxotrophs (his G-46, TA-1530) of <u>Salmonella typhimurium</u>, and (2) a diploid strain (D-3) of <u>Saccharomyces cerevisiae</u>. The induction of reverse mutation was determined with the <u>Salmonella</u>; mitotic recombination was determined with yeast. Chemicals were evaluated directly by <u>in vitro</u> bacterial and yeast studies prior to, or concurrent with, the studies in



mice. Only animals on the subacute studies were not fed the evening prior to compound administration. The Salmonella were carried in tryptone yeast extract gel, transferred weekly. They were transferred to tryptone yeast extract broth 48 hours before use: they were transferred a second time from broth to broth 24 hours prior to use, and again 8 hours before use. The mouse inoculum was prepared by transferring 4 ml of the 8-hour broth culture to 50 ml broth bottles which had been prewarmed at 37°C. Exponential log-phase organisms were inoculated intraperitoneally into the mice approximately 2-1/2 hours later when the appropriate density indicating 3.0 imes 10 $^8$ cells/ml was reached. The Saccharomyces was carried in yeast complete agar. The inoculum was prepared by harvesting the organisms from the surface of the plates with sterile saline. The cells were washed three times with sterile -saline and suspended in a concentration of 5.0  $\times$  10 $^8$  cells/ml. Two ml of the suspension was inoculated into each mouse intraperitoneally. Total plate counts on Salmonella were on tryptone yeast extract and for Saccharomyces on yeast complete medium.

## a. Acute study

Three dosage levels (usage, intermediate [determined as discussed previously], and LD $_5$ ) were administered orally by intubation to ten mice. Positive controls and negative vehicle controls were included in each study. All animals received 2 ml of the indicator organism intraperitoneally. Each ml contained 3.0 x  $10^8$  cells for Salmonella and 5.0 x  $10^8$  cells for Saccharomyces. Three hours later, each animal was killed and 2 ml of sterile saline was introduced intraperitoneally. As much fluid as possible was then aseptically removed from the peritoneal cavity. Dilution blanks for bacteria containing 4.5 ml of serile saline were prepared in advance. Tenfold serial



dilutions were made of each peritoneal exudate (0.5 ml exudate + 4.5 ml saline) yielding a concentration series from  $10^{\circ}$  (undiluted peritoneal exudate) through  $10^{-7}$ . For enumeration of total bacterial counts, the  $10^{-6}$  and  $10^{-7}$  dilutions were plated on tryptone yeast extract agar, 3 plates/sample, 0.2 ml sample/ plate. Each sample was spread over the surface of the plate using a bent glass rod immersed in 95% ethanol and flamed just prior to use. In plating for the total mutant counts on minimal agar, the  $10^{0}$  dilution was used, 0.2 ml being plated on each of 5 plates. The plating procedure was identical to that followed for the tryptone yeast extract agar plates. All plates were incubated at 37°C, tryptone yeast extract agar plates for 18 hours and minimal agar plates for 40 hours. For yeast mitotic recombination, dilution blanks containing 4.5 -mal of sterile saline were prepared in advance. Tenfold serial dilutions were made of each sample yielding a series from  $10^{0}$  to  $10^{-5}$ . Samples of 0.1 ml of the  $10^{-5}$ ,  $10^{-4}$ , and  $10^{-3}$  dilutions were removed and plated on complete medium (10 plates each). All plates were incubated at 30°C for 40 hours. The 10<sup>-5</sup> dilutions were used to determine total populations and the  $10^{-4}$  and  $10^{-3}$  plates were examined after an additional 40 hours at 4°C for red sectors indicating a mutation. Bacterial scoring was calculated as follows:

Total mutants on 5 plates x appropriate exponent = CFU/mI (CFU is Colony Forming Units) of sample plated CFU/ml x one/dilution factor  $(10^{0} - 10^{-7}) = CFU/mI$  in undiluted exudate. The mutation frequency (MF) calculated for each sample was:

MF = total mutant cells total population

 $MFt/MFc = \frac{MF \text{ of experimental sample}}{MF \text{ of control sample}}$ 

(MFt/MFc = 1.00 for control sample)



Yeast mitotic recombinants (presumptive ade 2, his 8 homozygotes) were seen as red colonies or as red sectors on a normally white yeast colony. The plates (from  $10^{-9}$  and  $10^{-3}$  dilutions) were scanned under the 10X lens of a dissecting scope to enumerate the red colonies and sectors. Population determinations were made from the  $10^{-5}$  dilution plates. A recombinant frequency (RF) was calculated:

# RF = total recombinants counted total number colonies screened

#### Subacute study

Similar groups of animals at each dose level received five oral doses of the test compound 24 hours apart. Within 30 minutes after the last dosing, the animals were inoculated with the test organism and handled in the same fashion as those in the acute study.

#### In vitro study

Cultures of <u>S. typhimurium</u> histidine auxotrophs

(G-46 and TA-1530) were plated on appropriate media. The test compound was then added to the plate, either in the form of a microdrop of solution (0.01 to 0.25 ml) applied to a small filter paper disc resting on the agar or a small crystal applied directly to the agar. Tenfold serial dilutions of the culture were employed and plated so as not to miss the optimum cell density for mutant growth.

Mutant colonies were observed and scored. Strain D-3 <u>Saccharomyces</u> cells at proper dilutions were shaken with the test compound, diluted, and plated at 50% survival level or above (see HMA Supplementary Materials and Methods). Red sectors were then scored and the frequency calculated after suitable incubation.

Negative and positive controls were run concurrently. The positive control was EMS for <u>Salmonella</u> and <u>Saccharomyces</u>. The <u>in vitro Salmonella</u> tests were reported



as (+) or (-) or questionable; the <u>in vitro Saccharomyces</u> tests were reported as sample concentrations, percent survival, and recombinants/ $10^5$  survivors. For the <u>Saccharomyces</u> a 50% survival level, e.g., an arbitrary 5.0% w/v test level, was used when no LD<sub>50</sub> was determinable.

#### 2. Cytogenetic Studies

#### In vivo study

Ten to twelve week old, male, albino rats obtained from a closed colony (random-bred) were used. A total of 59 animals in the acute study and 18 animals in the subacute study was used, as illustrated in the following protocol.

# Number of Animals Used

#### Acute Study

<u>Treatment</u>	Time Killed After Administration			
	<u>6 Hours</u>	24 Hours	48 Hours	
High Level	5	5	<sup>'</sup> 5	
Intermediate Level	5	5	5	
Low Level	5	5	5	
Positive Control	0	0	5	
Negative Control	3	3	3	

### Subacute Study

Five doses 24 hours apart; animals killed 6 hours after last dose.

Treatment	Killed After Administration
High Level	5
Intermediate Level	5 ·
Low Level	5
Negative Control	3

All animals were dosed by gastric intubation.

Four hours after the last compound administration, and two hours prior to killing, each animal was given 4 mg/kg of colcemid intra-



peritoneally in order to arrest the bone narrow cells in C-mitosis. Animals were killed by using CO<sub>2</sub>, and the adhering muscle and epiphysis of one femur were removed. The marrow "plug" was removed with a tuberculin syringe and an 18 gauge needle, aspirated into 5 ml of Hanks' balanced salt solution (BSS) in a test tube and capped. The specimens were centrifuged at 1,500 RPM in a table-top centrifuge for 5 minutes, decanted, and 2 ml of hypotonic 0.5% KCl solution was added with gentle agitation to resuspended the cells. The specimens were then placed in a 37°C water bath for 20 minutes in order to swell the cells. Following centrifugation for 5 minutes at 1,500 RPM, the supernatant was decanted and 2 ml of fixative (3:1 absolute methanol:glacial acetic acid) was added. The cells were resuspended in the fixative with gentle agitation, capped, and placed at 4°C for 30 minutes. The specimens were again centrifuged, decanted, 2 ml of prepared fixative was added, and the cells were resuspended and placed at 4°C overnight.

The following day the specimens were again centrifuged, decanted and 0.3 - 0.6 ml of freshly prepared fixative was added to obtain a suitable density. The cells were resuspended and 2 - 3 drops of the suspension were allowed to drop onto a clean, dry slide held at 15° from the horizontal. As the suspension flowed to the edge of the slide, it was ignited by an alcohol burner and allowed to flame. Following ignition, the slides were allowed to dry at room temperature overnight. Duplicate slides were prepared. The slides were stained using a 5% Giemsa solution (Giemsa buffer pH 7.2) for 20 minutes, rinsed in acetone, I:I acetone:xylene, and placed in fresh xylene for 30 minutes. The slides were then mounted using Permount (Fisher Scientific) and 24 x 50 mm coverglasses. The coverglasses were selected to be 0.17 mm  $\pm$  0.005 mm in thickness by use of a coverglass micrometer. The preparations



were examined using Leitz Ortholux I & II microscopes with brightfield optics and xenon light sources. These specimens were scanned with 10X and 24X objectives and suitable metaphase spreads that were countable were then examined critically using 40X, 63X or 100X oil immersion flatfield apochromatic objectives. Oculars were either 12X or 16X widefield periplanatics and the tube magnification either 1X or 1.25X. The filters used were either a didymium (BG20) or a Schott IL570 mu interference filter.

The chromosomes of each cell were counted and only diploid cells were analyzed. They were scored for chromatid gaps and breaks, chromosome gaps and breaks, reunions, cells with greater than ten aberrations, polyploidy, pulverization, and any other chromosomal aberrations which were observed. They were recorded on the currently used forms and expressed as percentages on the summary sheets. Fifty metaphase spreads were scored per animal. Mitotic indices were obtained by counting at least 500 cells and the ratio of the number of cells in mitosis/the number of cells observed was expressed as the mitotic index.

Positive controls in the acute study consisted of animals which had been given the known mutagen Triethylene Melamine (TEM) administered intraperitoneally at a level of 0.30 mg/kg. Negative controls on the acute and subacute studies consisted of the vehicle in which the compound was administered. The dosage levels, solvents and toxicity data are included in the Results and Discussion Section of the report.

# b. <u>In vitro</u> study

Human embryonic lung cultures (WI-38) which were negative for adventitious agents (viruses, mycoplasma) which may interfere



were used. These cells were employed at passage level 19. The cells had been transferred using 0.025% trypsin and planted in 32 oz. prescription bottles containing 40 ml of tissue culture medium. When growth was approximately 95% confluent the cells were removed from the glass using tryps:n, centrifuged, and frozen in tissue culture medium containing dimethyl sulfoxide (DMSO). Cells were frozen in vials in the vapor phase of liquid nitrogen at a concentration of 2  $\times$   $10^6$  cells/ml. When needed, the vials were removed from liquid nitrogen, quick-thawed in a 37°C water bath, washed free of DMS(), suspended in tissue culture medium (minimal essential medium [MEM] plus 1% glutamine, 200 units/ml of penicillin and 200 μg/ml of streptomycin and 15% fetal calf serum) and planted in milk dilution bottles at a concentration of 5 x  $10^5$  cells/ml. The test compound was added at three dose levels using three bottles for each level, 24 hours after planting. The dose levels required a preliminary determination of a tissue culture toxicity. This was accomplished by adding logarithmic doses of the compound in saline to a series of tubes containing 5  $\times$  10<sup>5</sup> cells/ml which were almost confluent. The cells were examined at 24, 48, and 72 hours. Any cytopathic effect (CPE) or inhibition of mitoses was scored as toxicity. Five more closely spaced dose levels were employed within the two logarithmic dosages, the higher of which showed toxicity and the lower no effect. The solvents used and the range finding data are presented in the toxicity data report under Results and Discussion. The dose level below the lowest toxic level was employed as the high level. Logarithmic dose levels were employed for the medium and low levels.

Cells were incubated at 37°C and examined twice daily to determine when an adequate number of mitoses were present. Cells were harvested by shaking when sufficient mitoses were observed, usually 24 - 48



hours after planting, centrifuged, and fixed in absolute methanol:glacial acetic acid (3:1) for 30 minutes.

The specimens were centrifuged, decanted, and suspended in acetic acid-orcein stain (2.0%) and a drop of suspension placed on a clean cry slide. Selected coverglasses 0.17 mm in thickness were placed on the suspension and the excess stain gently expressed from the slide. The coverglasses were sealed with clear nail polish and examined immediately.

The microscopes, objectives, oculars, filters and light sources were enumerated under the metaphase description. Positive controls used were TEM (at a concentration of 0.1 mcg/ml dissolved in saline) and negative controls which consisted of the vehicle in which the test compound was dissolved, which was 0.85% saline. Data were reported on forms currently used and expressed as percentages on the anaphase summary sheets.

#### Dominant Lethal Assay

In this test, male and female random bred rats from a closed colony were employed. These animals were 10-12 weeks old at the time of use. Ten male rats were assigned to each of 5 groups; 3 dose levels selected as described above, a positive control (triethylene melamine) (TEM) and a negative control (solvent only). The positive control was administered intraperitoneally. Administration of the test compound was orally by intubation in both the acute study (1 dose) and in the subacute study (1 dose per day for 5 days). Following treatment, the males were sequentially mated to 2 females per week for 8 weeks (7 weeks in the subacute study). Two virgin female rats were housed with a male for 5 days (Monday through friday). These two females were removed and housed in a cage until killed. The male was rested on Saturday and Sunday and two new females introduced to the cage on



Monday. It has been our experience that conception has taken place in more than 90% of the females by Friday and that the two day rest is beneficial to the male as regards subsequent weekly matings. Females were killed using CO<sub>2</sub> at 14 days after separating from the male, and at necropsy the uterus was examined for deciduomata (early deaths), late fetal deaths and total implantations.

Sufficient animals were provided in our experimental design to accommodate for any reduction in the number of conceptions. Each male was mated with two females per week, and this provided for an adequate number of implantations per group per week (200 minimum) for negative controls, even if there was a fourfold reduction in fertility of implantations. Results were analyzed according to the statistical procedures described in Supplementary Materials and Methods. Corpora lutea, early fetal deaths, late fetal deaths and total implantations per uterine horn were recorded on the raw data sheets, which are submitted separately.

- Supplementary Materials and Methods
  - Host-Mediated Assay <u>In Vitro</u> and Formulae
    - a. Bacterial in vitro plate tests

This method has been published by Ames: The Detection of Chemical Mutagens with Enteric Bacteria, in <u>Chemical Mutagens</u>; <u>Principles and Methods for Their Detection</u>, Vol. 1, Chapter 9, pp. 267-282, A. Hollaender, Editor, Plenum Press, New York (1971).

- b. <u>In vitro</u> for mitatic recombination
- (1) Strain D-3 was grown to stationary phase on complete medium agar plates at 30°C (3-4 days). Cells were rinsed from the plates and washed twice in saline and cell concentration determined spectro-



photometrically. (A standard curve previously determined for colony forming
units versus % transmittance at 545 mu was easily used.)

- (2) Cells from the concentration suspension were diluted appropriately into 0.067 M "hosphate buffer pK 7.2 to provide  $5 \times 10^7$  cells/ml in a total of 25 ml.
- 4 hours at 30°C, with shaking, at concentrations which permitted determination of the 50% survival level. Then, if not included in the first experiment, the compound was tested again only at the 50% survival level. If 50% survival level could not be determined, the arbitrary test level of 5% w/v was used.
- plated on complete agar medium for determination of total population and red sectors. Total surviving population was conveniently measured on plates of  $10^{-4}$  and  $10^{-5}$  dilutions using 0.2 ml per plate (5 plates), and sectors determined on plates of  $10^{-3}$  and  $10^{-4}$  dilutions using 0.2 ml per plate (5 plates). Plates were incubated for 2 days at 30°C followed by a holding period of 2 days at 4°C to promote color development with limited enlargement of the colonies. Red sectors were scored by systematically scanning the plates with a dissecting microscope at 10X magnification.
- (5) The frequency of red sectors can then be calculated and may be expressed conveniently as sectors per  $10^5$  survivors for comparison with untreated controls.
- (6) Ethyl Methane Sulfonate (EMS) was employed as the positive control in both <u>in vitro</u> systems.
  - c. Minimal medium (bacteria): Spizizen's Minimal Medium:



### 4X Salt Solution:

(NH<sub>A</sub>) SO<sub>A</sub>

8.0 gm

K2HPOA

56.0 gm

KH2PO4

24.0 gm

Na Citrate

4.0 gm

Mg 50,

0.8 gm

Biotin

0.004 gm

H<sub>2</sub>0

qs to 1 liter

Sterilize by autoclaving (121°C/15 min.)

#### Medium:

4X Salt Solution

:250 ml

5.0% Glucose (sterile)

:100 ml (If histidine is added at concentration of 30 mg/liter, this becomes a complete bacterial

medium.)

1.5% Bacto-agar (sterile)

:650 mT

d. Complete medium (bacteria):

Bacto-Tryptone

1.0 gm

Yeast-Extract -

0.5 gm

Bacto-Agar

2.0 gm

Distilled H<sub>2</sub>O

100.0 ml

Sterilize by autoclaving (121°C for 15 minutes).

Complete medium (yeast): e.

 $KH_2PO_A$ 

1.5 gm

MgSO<sub>4</sub>

0.5 gm

(NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>

4.5 gm



Peptone 3.5 gm Yeast-Extract 5.0 gm Glucose 20.0 gm Agar 20.0 gm Distilled  $\rm H_2O$  1000.0 ml

Sterilize by autoclaving (121°C for 15 minutes).

 Cytogenetics <u>In Vitro</u> Preparation of Anaphase Chromosomes (from Nichols, 1970)

"Anaphase preparations may be made by several methods. One convenient approach is to grow cells directly on coverslips in petri dishes. With human fibroblasts 400,000 cells added to a 22 x 44 mm coverslip in a 50 mm petri dish grown in a 5% CO2 atmosphere in air has proved very satisfactory. When adequate numbers of mitoses are visualized directly utilizing an inverted microscope (usually 48 to 92 hours after planting) the coverslip is transferred to absolute ethanol for 15 minutes for fixation. They are then stained with any one of a number of sultable stains (Fuelgen, May-Grunwald-Giemse, orcein) and attached to a slide with mounting media for evaluation. Anaphase preparations may also be prepared on cells grown in suspension or cells from a monolayer that have been put into suspension. In this instance the cells are centrifuged and fixed with the squash fixative. They are then suspended in the stain and a drop of the suspension put on the slide and covered with a coverslip. However, in this case, only the excess stain is gently expressed from under the coverslip and no squashing is carried out. In anaphase preparations no pretreatment with colchicine or hypotonic expansion is used and no technique for spreading the cells is used, so that the spindle and normal relationships of the chromosomes are not disturbed."



- 3. Statistical Analyses of Dominant Lethal Studies

  The following statistical analyses were employed as a
  means of analyzing the results of the dominant lethal studies.
  - a. The fertility index

The number of pregnant females/number of mated females with the chi-square was used to compare each treatment to the control. Armitage's trend was used for linear proportions to test whether the fertility index was linearly related to arithmetic or log dose.

Total number of implantations

The t-test was used to determine significant differences between average number of implantations per pregnant female for each treatment compared to the control. Regression techniques were used to determine whether the average number of implantations per female was related to the arithmetic or log dose.

c. Total number of corpora <u>lutea</u>

The t-test was used to determine significant differences between average number of <u>corpora lutea</u> per pregnant female for each treatment compared to the control.

d. Preimplantation losses

Preimplantation losses were computed for each female by subtracting the number of implantations from the number of <u>corpora lutea</u>.

Freeman-Tukey transformation was used on the preimplantation losses for each female and then the t-test was used to compare each treatment to control. Regression technique was used to determine whether the average number of pre-implantation losses per female was related to the arithmetic or log dose.



e. Dead implants

Dead implants were treated the same as pre-

implantation losses.

f. One or more dead implants

The proportion of females with one or more dead implants was computed, each treatment compared to control by chi-square test and Armitage's trend used for linear proportions to see if proportions were linearly related to either arithmetic or log dose. Also, probit regression analysis was used to determine whether the probit of the proportions was related to log dose.

g. Two or more dead implants

The proportion of females with two or more dead implants computed was treated same as above (f).

h. Dead implants per total implants

Dead implants per total implants were computed for each female and used Freeman-Tukey arc-sine transformation on data for each female; then used t-test to compare each treatment to control.

Historical control data was compiled on a continuous basis as studies were completed. In addition to comparing each treatment to control, as outlined above, each treatment was compared to a historical control.

In order to take variation between males into account, a nested model was used. An analysis of across weeks is also provided.

In addition to these tests, the distribution forms of the various parameters were tested in order to evaluate the appropriateness of some of the tests being used. Certain correlations between parameters may exist and were examined as one step to determine the appropriateness of models. If necessary, alternate test methods were implemented.



The results are presented in tabular form with the addition of historical control information. In addition to these tables, a written report of all findings is provided. As information became available from the on-going investigation of these data, it was reported and suggestions included for changes to the methods of analysis. The statistical reports give the level of significance using both a one-tailed and two-tailed test. Finally, a summary sheet for each study is provided.



Yisk = A + di + cis + cisk

ن المراجع المار عراسان Males within each group

🕨 👣 🗻 - Females within Males within Groups

SUMPTIONS

$$= \alpha_1 + \omega_2 = 0 \quad , \quad \text{cis} \quad \neg \text{nid}(o_1 o_2^2) \quad ,$$

hales are randomly drawn from infinite population

<u> </u>	<u>d.f.</u>	<u> </u>	MS	E(MS)	,
TOTAL	.39	55% (Yijk - 9)2			T
GROUPS MALES		20 E ( G G)2	S.~	6 + 262 + 2024	
W. HIN GROUPS	.18	= azz (4"ii - 4"i. )2		53+2Q2	1 ( ) ( ) ( ) ( ) ( ) ( )
LEMAINDER	ا ود ا	** ** ** ** ** ** ** ** ** ** ** ** **		0	

#### E. References

- Host-Mediated Assay
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## F. Abbreviations

- l. mu = micron
- mcg = ug = microgram
- g \* gram
- 4. kg = kilogram
- 5. ml = milliliter
- 6. rpm = revolutions per minute
- °C = degrees centigrade
- pH = power of the hydrogen ion concentration to the base 10
- 9. M = molar solution
- 10. conc. = concentration
- 11. MTD = maximum tolerated dosage = High = LD<sub>S</sub> if determined or else exceedingly high dose, such as 5 g/kg
- 12. INT \* intermediate = medium level
- 13. USE = usage level if known = low level
- 14. BSS = balanced salt solution
- 15. C-metaphase = cells arrested in metaphase, using colchine or colcemid
- 16. LD<sub>50</sub> = that dosage which produced 50% mortality in the group of animals treated
- 17. LD<sub>5</sub> = that dosage which produced 5% mortality in the group of animals treated
- 18. NC = negative control
- PC = positive control
- AU = acute usage level (low level)
- 21. AI = acute intermediate level (medium level).
- 22. AMTD = acute maximum tolerated dose level (LD<sub>5</sub> level, high level)



- 23. SAU = subacute usage level (low level)
- 24. SAI = subacute intermediate level (medium level)
- 25. SA LD<sub>5</sub> = subacute LD<sub>5</sub> level (MTD level, high level)
- 26. CO2 = carbon dioxide
- 27. DMN = Dimethyl nitrosamine
- 28. EMS = Ethyl methane sulfonate
- 29. TEM = Triethylene melamine
- 30. DMSO = Dimethyl sulfoxide
- 31. MEM = minimal essential medium (Eagle's)
- 32. CPE = cytopathic effect
- 33. his = histidine marker
- 34. D-3 = mitotic recombinant strain of Saccharomyces
- 35. mf = mean mutant frequency
- 36. MFt/MFc = mean mutant frequency of the test compound group compared to mean mutant frequency of the negative control group
- 37. CFU = colony forming units
- 38. WI-38 = code name for a strain of human embryonic lung tissue culture cells
- 39. Rec x  $10^5$  = mitotic recombinants x  $10^5$
- 40. Mean B/A = mean frequency
- 41. tot. scr. = total scored
- 42. tot. = total
- 43. X<sup>2</sup> = a test of variation in the data from the computed regression line tested in these studies at the 5% level
- 44. Aber. \* aberrations
- 45. Frag. = fragment
- 46. HMA = host-mediated assay

